Differential roles of ventral pallidum subregions during cocaine self-administration behaviors

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Abbreviations: NAcc – nucleus accumbens; VP – ventral pallidum; VPvm - ventromedial ventral pallidum; VPdl - dorsolateral ventral pallidum; S^D - discriminative stimulus; FR - firing rate; BLA - basolateral amygdala; VTA - ventral tegmental area; MDT - mediodorsal thalamus; PETH - perievent time histogram

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Abstract

The ventral pallidum (VP) is necessary for drug-seeking behavior. VP contains ventromedial (VPvm) and dorsolateral (VPdl) subregions which receive projections from the nucleus accumbens shell and core, respectively. To date, no study has investigated the behavioral functions of the VPdl and VPvm subregions. To address this issue, we investigated whether changes in firing rate (FR) differed between VP subregions during four events: approaching toward, responding on, or retreating away from a cocaine-reinforced operandum, and a cocaine-associated cue. Baseline FR and waveform characteristics did not differ between subregions. VPdl neurons exhibited a greater change in FR compared to VPvm neurons during approaches toward, as well as responses on, the cocaine-reinforced operandum. VPdl neurons were more likely to exhibit a similar change in FR (direction and magnitude) during approach and response than VPvm neurons. In contrast, VPvm firing patterns were heterogeneous, changing FRs during approach or response alone, or both. VP neurons did not discriminate cued behaviors from uncued behaviors. No differences were found between subregions during the retreat and no VP neurons exhibited patterned changes in FR in response to the cocaine-associated cue. The stronger, sustained FR changes of VPdl neurons during approach and response may implicate VPdl in the processing of drug-seeking and drug-taking behavior via projections to subthalamic nucleus and substantia nigra pars reticulata. In contrast, heterogeneous firing patterns of VPvm neurons may implicate VPvm in facilitating mesocortical structures with information related to the sequence of
behaviors predicting cocaine self-infusions via projections to mediodorsal thalamus and ventral tegmental area.
INTRODUCTION

First conceptualized by Heimer and colleagues (Heimer and Wilson, 1975; Heimer, 1978), the ventral pallidum (VP) was hypothesized to be a primary participant in the neuronal control of motivated behaviors (Mogenson et al. 1980). Over thirty subsequent years of research, VP has been shown to be critical for natural reward-seeking and drug-seeking behaviors. In rodent models of drug seeking, VP lesions or pharmacological challenges block morphine-induced sensitization (Johnson and Napier, 2000; Mickiewicz et al. 2009), drug-induced conditioned place preference (Gong et al. 1997; Rademacher et al. 2006; Dallimore et al. 2006), self-administration (Robledo and Koob, 1993), and reinstatement (McFarland and Kalivas, 2001; McFarland et al. 2004; Tang et al. 2005). In humans, post mortem analyses from cocaine abusers have shown alterations in VP neuropeptide expression (Frankel et al. 2008). However, akin to its primary afferent, the nucleus accumbens (NAcc), VP contains two “neurochemically distinct subterritories” that differ in their projection patterns (Zahm et al. 1996) and likely exhibit differential roles in drug-seeking behavior. Whether or not they do remains untested and is the subject of the present investigation.

The entire VP is characterized by dense substance P-immunoreactive fibers (Haber and Nauta, 1983; Groenewegen and Russchen, 1984; Haber et al. 1985; Zahm and Heimer, 1988; 1990; Zahm, 1989; Haber et al. 1990; Heimer et al. 1991; Groenewegen et al. 1993; Kuo and Chang, 1992; Kalivas et al. 1993;
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Zahm et al. 1996). The ventromedial subregion (VPvm) is demarcated by dense neurotensin-immunoreactive fibers and a lack of calbindin-d28k immunoreactivity (Zahm and Heimer, 1988, 1990; Zahm 1989; Zahm et al. 1996; Geisler and Zahm, 2006a). Conversely, the dorsolateral subregion (VPdl) is distinguished by dense calbindin-d28k immunoreactive fibers and a lack of neurotensin-immunoreactivity (Zahm et al. 1996; Riedel et al. 2002; Tripathi et al. 2010).

In addition to their distinct immunoreactivities, VPvm and VPdl subregions participate in independent neuronal circuits. The neurotensin-immunoreactive VPvm receives projections from the NAcc shell and projects predominantly to the mediodorsal thalamus (MDT) and ventral tegmental area (VTA) (Zahm and Heimer, 1988, 1990; Zahm, 1989; Zahm and Heimer, 1990; Heimer et al. 1991; Zahm and Brog, 1992; Groenewegen et al. 1993; Kuroda and Price, 1991; Kalivas et al. 1993; Zahm et al. 1996; Churchill et al. 1996; O'Donnell et al. 1997; Tripathi et al. 2010). In contrast, the calbindin-d28k immunoreactive VPdl receives projections from the NAcc core (which exhibits selective calbindin-d28k immunoreactivity compared to shell (Zahm and Brog, 1992)) and projects predominantly to the substantia nigra pars reticulate and subthalamic nucleus, and weakly to MDT (Zahm and Heimer, 1990; Groenewegen et al. 1993; Bell et al. 1995; Zahm et al. 1996; O'Donnell et al. 1997; Tripathi et al. 2010).

The discrete projection patterns of VPvm and VPdl suggest that the VP subregions differentially process behaviors within the ventral striatopallidal circuit. To the best of our knowledge no study has examined whether these subregions exhibit differential behavioral functions. The present study recorded single-units...
during cocaine self-administration from the VPvm and VPdl, as delineated by substance P, calbindin-d28k, and neurotensin immunohistochemistry. First, we extend our preliminary characterization of VP single-units during cocaine self-administration (Root et al. 2010) by determining which behaviors were coincident with changes in firing rate (FR) during self-administration. Second, we examined whether changes in FR during distinct cocaine-seeking behaviors differed between VP subregions.

NAcc neurons exhibit changes in FR during distinct components of drug-seeking behaviors including approaching and responding on a cocaine-reinforced lever (Chang et al. 1994, 1997, 2000). However in our experience with a cocaine-reinforced lever press, rats often exhibit a uniform ambulatory behavior that obscures both offset of approach and onset of response, as well as offset of response and onset of retreat. Therefore, we utilized a well-characterized long distance vertical head movement as the operant (Pederson et al. 1997; Tang et al. 2007; Pawlak et al. 2010; Root et al. 2011) which allowed the ability to discriminate onsets and offsets of approach, response, and retreat behaviors during self-administration. Specifically, the placement of the operandum within the corner of the self-administration chamber precluded locomotion during the operant response (i.e., alternating limb movements; Root et al. 2011). In contrast, the preceding approach towards, and subsequent retreat away from the operandum, involved turning and ambulatory behaviors.

Prior subregional examinations of NAcc neurons during cocaine self-administration revealed that NAcc core neurons exhibit greater changes in FR
than neurons in the medial NAcc shell over two time periods: 1) prior to response completion and 2) following response completion (Ghitza et al. 2004, 2006; Hollander and Carelli, 2005; Fabbricatore et al. 2010). The epoch “prior to response completion” consists of two distinct behaviors, approaching toward and responding on the operandum, while the epoch “following response completion” partially consists of retreating away from the operandum. Utilizing the vertical head movement operant, we distinguished changes in FR during approaches toward the operandum, responses on the operandum, and retreats away from the operandum. In order to investigate functional compartmentalization in the VP, we analyzed whether changes in FR during approach, response, or retreat alone, as correlations between two of these behaviors’ changes in FR, differed between VP subregions. Given that the NAcc core projects to the VPdl while NAcc shell projects to the VPvm, it was predicted that VPdl neurons would exhibit greater changes in FR than VPvm neurons during approach and response (i.e. prior to response completion) as well as retreat behaviors (i.e. following response completion).

MATERIALS & METHODS

Subjects and surgery

Male Long–Evans rats (n = 25; 320–340g; Charles River, Wilmington, MA) were anesthetized with sodium pentobarbital (approximately 50 mg/kg, i.p.). Prior to surgery, the rats received injections of atropine methyl nitrate (10 mg/kg, i.p.) and penicillin G (75,000 U/0.25 ml, i.m.) to reduce the risk of pulmonary
edema and bacterial infection, respectively. To block post-surgical pain sensitivity, animals were injected subcutaneously with 0.25 ml bupivacaine HCl (0.25%) (Abbott Laboratories, North Chicago, IL) spread over 8 injection sites of the incisions on the head and neck. Anesthesia was maintained with periodic i.p. injections of ketamine hydrochloride (approximately 60 mg/kg, i.p.) and sodium pentobarbital (approximately 15 mg/kg, i.p.). Following catheter implantation into the right jugular vein, a pair of 2x4 arrays (Microwire Technologies, Heightstown, NJ) composed of Quad Teflon-coated stainless steel microwire electrodes (California Fine Wire, Grover Beach, CA) were implanted bilaterally into the VP (between ± 0.7 to -0.9 AP, ± 0.8 to ± 3.1 ML, ±8.0 DV; Paxinos and Watson, 1997) and secured with dental cement. The diameter of each uninsulated microwire tip was 50 µm and wires were separated by 300 - 400 µm (center-center). An insulated 0.01 inch ground wire, stripped 5 mm from the tip, was implanted 5.5 mm ventral from the skull. The micro connector strip (Omnetics, Minneapolis, MN) was oriented mediolaterally (ear to ear), anterior to the interaural line, which prevented the harness cabling from breaking photocells during the task. After surgery, rats were individually housed with access to food and water in the cocaine self-administration chambers to recover for at least one week. Animals lived in the self-administration chambers and were maintained at 320-350g body weight with ad lib access to water and approximately 12-24 g of lab chow (Rat Diet #5012, Purina Mills Nutrition International, St. Louis, MO) following daily self-administration sessions. Rats received infusions of heparinized saline every 25 minutes other than during self-administration
sessions to preserve catheter patency. Mild food restriction and heparinized saline injection procedures were also used to facilitate backward compatibility between NAcc core and shell datasets from our laboratory (Peoples et al. 1997; Ghitza et al. 2004; Fabbricatore et al. 2010). Protocols were performed in compliance with the Guide for the Care and Use of Laboratory Animals (NIH, Publications 865–23) and approved by the Institutional Animal Care and Use Committee, Rutgers University.

Electrophysiological procedures

Sixteen neural signals were led through four quad-channel operational amplifiers (TLC2264, Texas Instruments, Dallas, TX) inside a custom recording harness (MB Turnkey Designs, Hillsborough, NJ) and fluid/electronic swivel (CAY-675–24, Airflyte Electronics, Bayonne, NJ) to a preamplifier (MB Turnkey Designs, Hillsborough, NJ) that differentially amplified (10x) the signal on the recording electrode against another microwire that did not exhibit a single unit.

The signal was then band-pass filtered (450 Hz to 10 kHz; roll off 1.5 dB/octave at 1 kHz and -6 dB/octave at 11 kHz) and amplified 700x between rollovers (MB Turnkey Designs, Hillsborough, NJ). Using software and hardware of DataWave Technologies, electrical signals were sampled (50 kHz sampling frequency per wire) and stored for offline analysis.

One recording of all microwires per animal occurred between days 14 and 24 of self-administration training. The rationale for recording at this time point was as follows: First, we have previously demonstrated rapid phasic changes in
FR surrounding cocaine-reinforced responses during this time period (Root et al. 2010). Second, this time point follows the development of escalated drug intake (Root et al. 2011), posited to exemplify "addiction-like behavior" in rats (Ahmed and Koob, 1998; Deroche-Gamondet et al. 2004). Third, well-trained animals quickly load to stable asymptotic drug levels that exhibit little variability during the recording session, mitigating pharmacological differences between responses (Root et al. 2011).

Isolation and separation of individual neural waveforms from background noise and waveforms of other neurons recorded from the same microwire were conducted post-hoc using DataWave spike sorting and separation software. First, neural discharges were sorted using waveform parameters, including principal components one and two, valley voltage, peak voltage, voltages at user-defined time cursors, spike height, and peak voltage time. Scatter plots of any 2 waveform parameters were displayed in a window, with all windows' parameter combinations displayed on one screen simultaneously. Each point plotted on the scatter plot corresponded to one recorded waveform. Each cluster of dots represented similar waveforms, which were separated from other clusters by enclosing it within an elliptical window. All waveforms of the putative individual neuron during the entire session (6-10 hours) were then replayed on a computer-simulated oscilloscope in order to assess the stability of neural waveforms. Waveforms whose parameters did not remain stable were discarded. Second, an interspike interval histogram was constructed. If discharges occurred within the first 2 ms in the interspike interval, corresponding to a neuron's natural refractory
period, the recording was not considered that of a single neuron and discarded.

When more than a single population of neural waveforms appeared to have been recorded from a given wire, cross-correlation was used to confirm that the populations corresponded to distinct neurons. Specifically, if discharges occurred within the first 2 ms in the cross-correlation and both neurons contained 0 discharges within their interspike intervals, both neurons were considered independent single units. Neurons exhibiting signal-to-noise ratios less than 2:1 were discarded.

Head movement task

Long distance vertical head movements were selected as the operant. First, this operant facilitates rapid shaping as well as the ability to characterize certain parameters of the movement during water or cocaine self-administration, as our laboratory has done previously (Pederson et al. 1997; Tang et al. 2007; Pawlak et al. 2010; Root et al. 2011). Second, using this operant we have demonstrated the acquisition of skilled cocaine self-administration behavior (Root et al. 2011), hypothesized to occur in human drug users (Tiffany, 1990). Third, this apparatus measures operant movements, and readily distinguishes them from any stereotypical (e.g. cocaine-induced) movements that occur at 10 Hz (Root et al. 2011; Fowler et al. 2003, 2007; also see Movies 1 and 2) but are rarely emitted in the corner containing the apparatus.

The apparatus tracked vertical head movements with infrared-emitting diodes capable of transistor-transistor logic (HOA6299, Honeywell, Morristown,
Six photocells were arranged along a 50 degree arc over 69 mm in one corner of the self-administration chamber. The lowest of six photocell beams, photocell one, was positioned 13 mm above the floor. Photocell six was the highest photocell. For full apparatus details and several images see Root et al. (2011). For neuronal analysis during vertical head movements, timestamps of photocell beam breaks were collected using DataWave Technologies, which allowed a unique decimal code for simultaneous beam breaks (i.e. photocells beams 1 and 2 broken at the same time). Two simultaneous photocell beam breaks of consecutive photocells were considered as half steps, equally between two photocells. For example, simultaneously breaking photocells 2 and 3 was considered 2.5. For neuronal analyses during responses, movement onset was defined as a photocell beam break for which at least the next photocell beam break was 0.5, 1, or 1.5 larger than the previous photocell beam break and less than one second in duration from the original photocell beam break. Offset was defined as the last instance for which the previous photocell beam break was 0.5, 1, or 1.5 larger than the previous photocell beam break and less than one second in duration from the onset. For both behavioral and neuronal analyses, all photocell beam breaks were recorded for offline analysis of different movements (1 millisecond resolution). A criterion head movement was defined as a vertical head movement breaking photocells two, three, four, and five consecutively within one second (schematized in Figure 1B). The criterion head movement required at least a 43 mm movement (c.f. Tang et al. 2007; Pawlak et al. 2010),
which could start at or below the 2nd photocell beam and cross the fifth or sixth photocell.

Self-administration began daily with the onset of the house light. Self-administration sessions were controlled by custom programs written in MED-PC (MED-Associates, St. Albans, VT). At the beginning of the first day, vertical head movements in the "head movement corner" were shaped via delivery of cocaine infusion (0.24 mg/0.2 ml/7.5 sec inf) by the experimenter in the presence of a discriminative stimulus (SD) tone (3.5 kHz, 70 dB). During acquisition, the SD was sounded for 2 min or until a criterion head movement was made. If a criterion movement was made, three events occurred simultaneously: the SD was terminated, cocaine delivery began, and a 40 second time out period began. If a criterion movement was not made, the SD was terminated after 2 min and a 5 second time out period began. The SD was sounded again following the time out period. Criterion movements within the time out period were recorded but had no programmed consequence. Thus, the SD set the occasion for each cocaine-reinforced movement, as in to previous investigations of NAcc shell and core neurons during cocaine self-administration (Ghitza et al. 2004). Operant responses were readily shaped within the first day in most rats. The shaping process began with rewarding breaking photocell 2 for ten rewards, then rewarding breaking photocells 2 and 3 consecutively within one second for five rewards. Shaping continued with rewarding breaking photocells 2, 3, and 4 consecutively within one second for five rewards and lastly rewarding the final contingency of breaking photocells 2, 3, 4, and 5 within one second.
consecutively. Most rats self-administered at the final contingency for two additional days under the same $S^D$ duration, infusion dose, time out duration, and contingency.

Following acquisition (starting typically on day 4), animals began normal training. Normal training consisted of the same FR1 schedule of reinforcement during $S^D$ presentations. To maximize the amount of time the animal spent at asymptotic drug levels where electrophysiological analyses occurred (see Neuronal analysis), prior to the tenth self-infusion the $S^D$ was sounded for 30 sec and the time out was fixed to 5 sec. All subsequent time outs and $S^D$ durations were fixed to 30 sec with a dose of 0.06 mg/0.05 ml/1.875 sec inf in order to increase the number of events used in neuronal analyses. In all cases, when the rat satisfied the criteria for a criterion head movement while the $S^D$ was sounded, three events occurred simultaneously, the $S^D$ was terminated, cocaine delivery began, and the time out began. If the rat did not execute a criterion head movement during $S^D$ presentation, the $S^D$ was terminated after 30 sec and the time out began. Criterion head movements that occurred when the $S^D$ was off (time out) were recorded but had no programmed consequence. The present schedule of reinforcement allowed rats to attain a maximum instantaneous calculated drug level of 7.16 mg/kg, which was above the level self-administered by all rats. That is, each rat was able to self-administer its “preferred” level, or satiety threshold level of drug (Root et al. 2011). During training, self-administration sessions ended after 6 hours elapsed or 280 rewards were earned, whichever occurred first. During the recording session, the self-
administration session ended six hours following the tenth self-infusion of cocaine. Overnight between self-administration sessions, a rectangular 2 in x 2 in x 8 in Plexiglas block was fastened into the head movement corner to block extinction learning. Rats were never “primed”, i.e., noncontingently administered infusions of cocaine to initiate self-administration.

Histological procedures

At least one hour following the final self-administration session, animals were injected subcutaneously -18 h and -4 h prior to perfusion with 2 mg/kg of haloperidol in order to enhance subsequent immunohistochemical staining in the VP (Eggerman and Zahm, 1988; Zahm and Heimer, 1990; Geisler and Zahm, 2006a). At 0 h, animals were anesthetized with an overdose of sodium pentobarbital (150-200 mg/kg, i.p.). Anodal current (50 µA for 3 sec) was passed through each microwire to mark the tip location and the animal was perfused with saline followed by 4% paraformaldehyde. The brain was removed and fixed in a solution of 4% paraformaldehyde overnight. Subsequently, the brain was stored in a 30% sucrose-phosphate buffer solution at 4°C. Brains were coronally sliced at a thickness of 40 µm through VP and placed in phosphate buffer filled 10 mL net wells for immunohistochemistry.

All immunohistochemical steps were carried out under gentle agitation on a horizontal rotator (Laboratory-Line, Fisher, Pittsburgh, PA, USA). Free-floating sections were rinsed in 0.1 M phosphate buffer (pH 7.4), placed into 1% sodium borohydride for 15 min, thoroughly rinsed in 0.1 M phosphate buffer again,
pretreated with 0.1 M phosphate buffer containing 0.1% Triton X-100 and 3% normal goat serum for 1 h, and then transferred into a solution containing a primary antibody, either polyclonal anti-neurotensin (ImmunoStar, Inc., Hudson, WI; formerly DiaSorin Histochemical as well as INCSTAR; catalog number 20072) at a dilution of 1 : 6500, polyclonal anti-calbindin-d28k (ImmunoStar; catalog number 24427) at a dilution of 1 : 6000, or polyclonal anti-substance P (ImmunoStar catalog number 20064) at 1 : 6500 in 0.1 M phosphate buffer with 0.1% Triton X-100 and 3% normal goat serum overnight (see Table 1 and Antibody Characterization section for details regarding the primary antibodies used). Every first of three slices was prepared with anti-substance P, every second of three slices was prepared with anti-calbindin-d28k, and every third of three slices was prepared with anti-neurotensin.

The following day, after thorough rinsing in 0.1 M phosphate buffer with 0.1% Triton X-100, sections were placed in a solution containing biotinylated antibody against rabbit immunoglobulin (Vector Laboratories Inc., Burlingame, CA, USA), at a dilution of 1 : 200 in 0.1 M phosphate buffer with 0.1% Triton X-100 for 1 h. The sections were again rinsed in 0.1 M phosphate buffer with 0.1% Triton X-100 and immersed in a solution containing avidin-biotin-peroxidase complex (Vector; 1 : 200 in 0.1 M phosphate buffer containing 0.1% Triton X-100) for another hour. After thorough rinsing in 0.1 M phosphate buffer, a color reaction was developed by immersing the sections for 6 min in a solution of 0.01 M phosphate buffer containing 0.05% 3,3'-diaminobenzidine. Sections were mounted onto gelatin-coated slides, incubated in a solution of 5% potassium
ferrocyanide and 10% HCl to stain the iron deposits left at the marked microwire recording tip. Sections were then dehydrated through a graded series of alcohol, transferred into xylene, and coverslipped with Permount (Fisher, Pittsburgh, PA, USA). Sections were photographed, which were then enhanced by using “Hard Light” blend mode in Photoshop CS6 (Adobe Systems Incorporated, San Jose, CA).

Antibody characterization

The polyclonal anti-substance P antibody was raised in rabbit against synthetic substance P (RPKPPQQFFGLM) coupled to keyhole limpet hemocyanin with carbodiimide and shows no cross-reactivity with other brain peptides, including neurokinin A, neurokinin B, somatostatin, and neuropeptide K. According to the manufacturer, specific immunostaining with the antibody was completely abolished by preadsorption with substance P at a final concentration of 10 µg/ml. Several other laboratories have also reported the abolition of specific immunostaining with the antibody by preadsorption with substance P (Brog et al. 1993; Zahm et al. 1996; Weissner et al. 2006; Cantwell and Cassone, 2006; Kim et al. 2008).

The polyclonal anti-calbindin-d28k antibody was raised in rabbit against calbindin-d28k purified from bovine cerebellum and shows no cross-reactivity with other brain peptides, including calretinin, vasoactive intestinal polypeptide, somatostatin, substance P, neuropeptide Y (Buchan and Baimbridge, 1988; Meek et al. 2008). Western blot analysis of the antibody detected a single band.
of 28 kD (Buchan and Baimbridge, 1988; Bell et al. 2005; Meek et al. 2008).

According to the manufacturer and scientific literature (Buchan and Baimbridge, 1988; Conde et al. 1994; Zahm et al. 1996; Bouilleret et al. 2000), specific immunostaining with the antibody was completely abolished by preadsorption with calbindin-d28k, which according to Buchan and Baimbridge (1988) was at a final concentration of 1 nM.

The polyclonal anti-neurotensin antibody was raised in rabbit against synthetic peptide (human) neurotensin coupled to bovine thyroglobulin with glutaraldehyde and shows no cross-reactivity with other brain peptides, including neurokinin A, neurokinin B, somatostatin, and neuropeptide K (Immunostar; c.f. Geisler and Zahm, 2006b). According to the manufacturer and scientific literature (Zahm et al. 1996; Geisler and Zahm, 2006b; Zahm et al. 2011), specific immunostaining with the antibody was completely abolished by preadsorption with a final concentration of 10 µg/ml of neurotensin. No staining was observed when any of the three primary antibodies were omitted, which controlled for the secondary antiserum.

Designation of single-units to subregions of the VP

Designation of single-units to subregions of the VP was made by a scorer blind to changes in FR during self-administration. If all implanted microwire tracks were identified from their entry into cortex to their tips (blue spots marked by potassium ferrocyanide staining of iron deposits), microwire tip positions were subsequently histologically localized. If any of the implanted microwires could not
be identified, neural data from the animal were discarded. In order to be
designated into the VPvm, single-units belonging to potassium ferrocyanide-
stained tissue from microwire tips were localized: i) within substance P
immunoreactivity; ii) within neurotensin immunoreactivity; and iii) absent from
calbindin-d28k immunoreactivity (Zahm and Heimer, 1988, 1990; Zahm 1989;
Zahm et al. 1996). In order to be designated into the VPdl, single-units belonging
to potassium ferrocyanide-stained tissue from microwire tips were localized: i)
within substance P immunoreactivity; ii) within calbindin-d28k immunoreactivity;
iii) and absent from neurotensin immunoreactivity (Zahm et al. 1996; Riedel et al.
2002; Tripathi et al. 2010).

Training analysis

Outcome variables, e.g., criterion head movements, self-administered
mg/kg/day, etc., were analyzed as a function of fourteen training days using
repeated measures ANOVAs (PASW 18.0.0, SPSS, Chicago, IL). When the
assumption of sphericity was not confirmed, a Greenhouse-Geisser correction
was used which adjusts the degrees of freedom in the ANOVA to be more
conservative. Fourteen training days was selected because it was the time of the
earliest recording session. Alpha criterion for all tests was 0.05.

Video analysis

A video frame-counter (Thalner Electronics VC-436, Ann Arbor, MI),
connected to the neuronal clock (CTR05, Measurement Computing, Norton, MA),
time-stamped each frame in a videocassette recorder (JVC HR-DD84OU, Wayne, NJ), which was used to monitor and record additional behaviors (approach and retreat, Figure 1A and 1C, respectively) for off-line analysis (33 ms resolution).

Approach onset was determined by the start of a change in direction exhibited by the animal towards the photocell corner and culminated with the breaking of a photocell. Approach onset was typically a leftward or rightward head turn or a vertical head movement prior to head turning, consistent with the approach behaviors described by Chang and colleagues (1994, 1997, 2000). In the minority of cases for which animals initially faced the direction of the photocell corner while engaged in focused stereotypy, approach onset was determined as the start of alternating limb movements toward the photocell corner and culminated with the breaking of a photocell. Approach offset was determined by the first photocell break following the approach onset. Approaches and retreats in which the approach onset was less than 150 ms from a previous retreat offset (described below) were removed from analysis due to ambiguity in defining the offset of retreat and onset of approach. Approaches greater than three seconds in duration were excluded from analysis. Cued approaches were defined as approaches in which the $S^D$ was on at approach onset whereas uncued approaches were defined as approaches in which the $S^D$ was off at approach onset. Cued responses were defined as criterion movements in which the $S^D$ was on at response onset and uncued responses were defined as criterion movements that occurred when the $S^D$ was off. For cue analysis, an $S^D$ trial that
did not culminate in a self-administered cocaine infusion was defined as a “miss” and an $S^D$ trial that culminated in a self-administered cocaine infusion was defined as a “hit”.

Retreat onset was determined by the start of a change in direction exhibited by the animal away from the photocell corner. In the vast majority of cases, this was a leftward or rightward head turn but in a minority of cases was a backwards movement. The retreat offset was determined by the start of an additional change in direction exhibited by the animal or the start of a pause in locomotion of at least 66 ms (two video frames). For cases in which the animal's movement following retreat onset contained slight head bobbing or circular head movements during a continuous movement away from the photocell, these slight head movements were ignored and retreat offset was determined by the start of an overt additional change in direction exhibited by the animal. Retreats greater than five seconds in duration were excluded from analysis. Pump retreats were defined as retreats for which the pump was activated or deactivated within two seconds prior to or during a retreat whereas nonpump retreats were all other retreats. Four example approach, response, and retreat events are displayed in Movie File 1.

Neuronal analysis

Changes in FR were examined with respect to four events: cue, approach, response, and retreat. For each occurrence of an event, FR was calculated as number of spikes divided by the duration (sec) of that particular event. Then, all
of its occurrences within a session were grouped and an average FR was
determined. For the cue event, the objective was to minimize any contribution of
movement to the assessment of FR. The average FR was calculated between \( S^D \)
onset to \( S^D + 150 \) ms. For the approach event, the average FR was calculated
between approach onset and approach offset (detailed in Video Analysis
section). For the response event, the average FR was calculated between
criterion movement onset and offset (detailed in Head movement task section).

For the retreat event, the average FR was calculated between retreat onset and
retreat offset (detailed in Video Analysis section). Changes in FR were visualized
in Matlab (The Mathworks, Cambridge, MA) with raster and peri-event time
histograms (PETHs), the code of which was modified from Ma (2010).

Firing during the time points described above were considered potential
"signals" during behavior. These signals were compared to a baseline period for
standardized "signal:baseline" ratios using a 'B/(A+B)' formula, termed the
directional change in FR, where 'B' was the signal and 'A' was the baseline. For
the cue event, the baseline was the average FR during the 150 ms prior to cue
onset, as previously used (Ghitza et al. 2003). Pre-cue FR was not likely related
to any particular movement because behavior was “randomized” in that cues
were presented non-contingently. Similarly, cue-evoked movements were
prevented from influencing post-cue FR assessments because their onset
latency after \( S^D \) onset is >150 ms (Ghitza et al. 2003). For approach, response,
and retreat events the baseline was the average FR during the period from -6 to -3 sec prior to criterion movements, as previously used (Ghitza et al. 2004, 2006).
If within the baseline periods any photocell beam break, approach onset or offset, retreat onset or offset, or pump onset or offset occurred, that particular trial was omitted from calculating the baseline. The directional change in FR ranged between 0 and 1, with a value of 0.5 indicating no change from baseline, values less than 0.5 a decrease in FR from baseline, and values greater than 0.5 an increase in FR from baseline. Since VP neurons exhibit changes in FR surrounding cocaine-reinforced responses with heterogeneous decreases or increases in FR (Root et al. 2010), a second analysis was conducted on all events termed the absolute change in FR. The absolute change in FR was calculated using the formula \(| \text{directional change in FR} - 0.5 |\). The absolute change in FR ranged between 0 and 0.5 (where 0 = no change from baseline FR) and represents change in FR from baseline regardless of whether the change was a decrease or increase in FR. For all analyses (cues, responses, approaches, and retreats), events that occurred during the initial 10 rapidly-spaced “loading” self-infusions were excluded in order to minimize the influence of pharmacological differences between events.

To evaluate subregional differences in the magnitude of changes in FR between VPvm and VPdl neurons, Mann-Whitney U tests evaluated the absolute change in FR for cue, approach, response, and retreat events, as previously used (Ghitza et al. 2004, 2006; Fabbricatore et al. 2010). Mann-Whitney U tests were necessary because these data did not fit a normal distribution. If a significant difference was observed between subregions using the absolute change in FR during approach, response, or retreat, we examined whether the
populations exhibited differential increases/decreases in FR by using the Mann-Whitney U test on the directional changes in FR for that specific behavior. To examine differences in FR during cued and uncued behaviors, correlation coefficients from each subregion were computed using the directional change in FRs (e.g., the correlation coefficient of cued approach directional change in FR and uncued approach directional change in FR). To analyze differential relationships between subregions, correlation coefficients of directional changes in FR values were compared using a z-test as described by Morse (1999). To analyze subregional differences in the prevalence of recorded neurons in left versus right hemisphere, biphasic versus triphasic waveform profiles, and propensity to record from multisingle-unit microwires (single microwires that recorded more than 1 single-unit), Fisher's exact test was used. All statistics were used at the population level rather than single neuron level, in order to not violate the statistical assumption of independence.

RESULTS

Behavior

Prior to recordings, all rats exhibited both task and skill learning. With regard to task learning, reaction time in response to the SD decreased over days, $F(4.040, 96.970) = 99.468, p < 10^{-32}$ (Greenhouse-Geisser corrected; Figure 2A). Over training sessions, probability of self-infusion upon SD presentation, $F(4.424, 106.173) = 24.792, p < 10^{-14}$ (Greenhouse-Geisser corrected; Figure 2B), and
total drug intake (mg/kg), $F(4.980,119.530) = 8.647, p < 10^{-6}$ (Greenhouse-Geisser corrected; Figure 2C), increased. Animals escalated their intake of cocaine over days, decreasing the latency to reach the 10th self-infusion, $F(3.886,85.487) = 5.133, p < 0.001$ (Greenhouse-Geisser corrected; Figure 2D).

With regard to skill learning, the number of criterion movements increased over days, $F(4.948,118.759) = 33.18, p < 10^{-20}$ (Greenhouse-Geisser corrected; Figure 2E), while the number of inverse criterion movements (downward movements starting at or above photocell 5 and ending at photocell 2 or below, but without programmed consequence) did not change, $F(1.907, 45.762) = 2.116, p > 0.05$, Greenhouse-Geisser corrected. Over the last three days of training, the latency for the subsequent movement following a criterion movement was $19.26 \pm 2.77$, $17.80 \pm 2.97$, and $16.76 \pm 2.75$ sec, demonstrating that criterion movements were inconsistent with stereotypic head bobbing. The average velocity of criterion movements increased over days, $F(3.532, 84.775) = 9.480, p < 10^{-5}$ (Greenhouse-Geisser corrected; Figure 3A), as a function of decreased duration, $F(2.816, 67.583) = 20.747, p < 10^{-8}$ (Greenhouse-Geisser corrected; Figure 3B), but not distance, $F(13,312) = 27.116, p > 0.05$ (Greenhouse-Geisser corrected; Figure 3C). The probability of starting the criterion movement at the minimum requirement, photocell 2, increased over days, $F(6.193, 154.828) = 2.533, p < 0.05$ (Greenhouse-Geisser corrected; Figure 3D). The probability of ending a criterion movement at the minimum requirement, photocell 5, did not change, $F(4.686, 112.469) = 1.045, p > 0.05$ (Greenhouse-Geisser corrected; Figure 3E).
In summary, animals learned the self-administration task which consisted of a vertical head movement operant. Animals escalated their intake of drug, posited to exemplify "addiction-like behavior" in rats (Ahmed and Koob, 1998; Deroche-Gamonet et al. 2004). Although not required, animals became skilled and efficient in their use of the vertical head movement operant, consistent with our previous examination (Root et al. 2011), and hypothesized to occur in highly experienced human drug users (Tiffany, 1990).

Histological localization and baseline single-unit properties

Neurons were recorded typically on day 16 of training (average), ranging between days 14 and 26. Of the 400 microwires implanted within the basal forebrain of 25 rats, 202 microwires were localized to the VP. Of these, 38 microwires were localized to the VPdl (Figure 4A-I displays three examples; Figure 5 shows all microwires as blue triangles), recording 48 single-units. A total of 140 microwires were localized to the VPvm (Figure 4J-R displays three examples; Figure 5 shows all microwires as black circles), recording 171 single-units. Twenty-four microwires recording 24 single-units were removed due to histological localization to all three stains, which typically occurred at the most caudal, sublenticular aspects of VP. The amplitude and signal-to-noise ratio of VP neurons (Table 2) did not differ between subregions (all $|z| < 1.84$, $p > 0.05$). The prevalence of recording from multiple single-unit microwires, 9/38 (23.6%) of VPdl and 24/140 (17.14%) of VPvm microwires, did not differ between subregions (Fisher's exact test, $p > 0.05$). A minority of single-units exhibited an
initial positivity prior to the valley of the waveform (triphasic profile), 4/48 (8.33%) in VPdl and 20/171 (11.70%) in VPvm, the prevalence of which did not differ between subregions (Fisher's exact test, p > 0.05). The prevalence of single-units recorded from microwires within the right versus left hemisphere, with 20/48 (41.67%) of VPdl and 88/171 (51.46%) of VPvm single-units recorded from the right hemisphere, did not differ between subregions (Fisher's exact test, p > 0.05). The cue baseline FRs (Table 2) and behavior baseline FRs (Table 2) did not differ between subregions (all |z| < 0.28, p > 0.05). Overall, these results demonstrate that waveforms and baseline FRs of recorded neurons were not statistically different between VP subregions and hemispheres.

Analytical strategy

During cocaine self-administration, subsets of NAcc neurons exhibit changes in FR during prior to response completion, following response completion, or both time periods (Hollander and Carelli, 2005; Ghitza et al. 2004, 2006; Fabbricatore et al. 2010). In the present study, in which we have specifically isolated approach, response, and retreat behaviors, 85.42% of VPdl and 76.02% of VPvm neurons exhibited at least a 20% change in FR during one, or any combination of, two or three, of these behaviors (Table 4). See Table 3 for average ± SEM number of approach, response, and retreat events per animal and their average ± SEM durations (ms) used in electrophysiological analyses. VP neurons distributed themselves into smaller subsets of neurons across the different components of the task. Therefore, the following results are organized
by neurons that exhibited changes in FR during one behavior only, two behaviors, and finally all three behaviors. Furthermore, we tested whether changes in FR during these behaviors, as well as the correlations of changes in FR between different behaviors, differed between VPdl and VPvm subregions.

The major differences between VP subregions occurred during approach and response components of self-administration. First, VPdl neurons exhibited a larger change in FR than VPvm neurons during approach or response. In addition, changes in FR by VPdl neurons typically began during the approach and carried through the response in the same direction with similar or greater magnitude. In contrast, VPvm neurons exhibited significantly weaker changes in FR during approach and response, as well as a significantly weaker relationship of changes in FR between approach and response.

Changes in FR during one of three cocaine-seeking behaviors

The three behaviors investigated here (approach, response, and retreat), are important for different reasons. The approach can be considered a cocaine-seeking behavior whereas the response can be considered a cocaine-taking behavior. In contrast, the retreat is a behavior that might reflect the anticipated reward (cocaine infusion) for the immediately preceding completion of cocaine-seeking behaviors (approach, response) and/or a behavior that leads to the location of focused stereotypy. Woodward and colleagues (Chang et al 1994, 1997, 2000) have shown NAcc neurons exhibit changes in FR during distinct behaviors related to cocaine self-administration. We first investigated whether VP
neurons discriminated the components of cocaine self-administration (approach, response, or retreat) and whether changes in FR during these behaviors differed between VP subregions. First, discrimination of different behaviors by VP neurons would demonstrate that the behaviorally-coincident firing patterns observed in NAcc neurons during cocaine self-administration (Chang et al. 1994, 1997, 2000) are also observed in VP. Second, differential changes in FR between VP subregions during individual components of cocaine self-administration would elucidate the individual contributions of the ventral striatopallidal subcircuits.

Subpopulations of neurons that exhibited changes in FR during only the approach towards the photocell corner were observed in both VP subregions (Table 4). However, as a population during the approach, neurons of the VPdl exhibited a significantly larger absolute change in FR than neurons of the VPvm ($z = -3.30$, $p < 0.001$) (Figure 6A). Two examples of approach-related firing patterns are shown in Figure 6B (VPdl neuron) and Figure 6C (VPvm neuron). In each case, changes in FR occur at the onset of approach (Figure 6B (left), 6C (left)) and exhibit no change in FR during response (Figure 6B (center), 6C (center)) or retreat (Figure 6B (right), 6C (right)). Subregional differences were not observed for the approach when examining directional changes in FR, $z = -0.85$, $p > 0.05$, suggesting approach-related firing was heterogeneous in direction for both subregions.

To examine the influence of the S^D upon approach-related firing patterns, approaches were sorted into cued and uncued trials. Between subregions, the
VPdl exhibited larger cued (Figure 6D) and uncued approach (Figure 6E) absolute changes in FR than the VPvm (cued z = -2.90, p < 0.01; uncued z = -2.26, p < 0.05). Within the VPdl alone, a significant correlation was observed between cued and uncued approach directional changes in FR, VPdl $r = 0.69$, p $< 10^{-7}$ (Figure 6F). Within the VPvm alone, cued and uncued approach directional changes in FR were also significantly correlated, VPvm $r = 0.73$, p $< 10^{-28}$ (Figure 6G). The correlation coefficients did not differ between subregions, $z = -0.45$, p $> 0.05$. In other words, within each subregion most neurons did not discriminate changes in FR between cued approach and uncued approach conditions. Two examples of similar firing patterns between cued and uncued approaches are displayed in Figure 6H for a VPdl neuron and Figure 6I for a VPvm neuron. On the rare occasion in which a difference existed between cued and uncued approach conditions, such neurons were located in the VPvm. Thus, it appears that the $S^D$ during cocaine self-administration has little to no influence upon approach-related firing patterns of individual neurons from either VP subregion.

The most common changes in FR of VP neurons occurred during only the response (criterion movement; Table 4). As a population during the response, neurons of the VPdl exhibited a significantly larger absolute change in FR than neurons of the VPvm (z = -2.78, p < 0.01) (Figure 7A). Two examples of response-related firing patterns are shown in Figure 7B (VPdl neuron) and Figure 7C (VPvm neuron). In both cases, FRs were altered during the response (Figure 7B (center), 7C (center) but not the retreat (Figure 7B (right), 7C (right)). The VPvm neuron was not altered during the approach (Figure 7C (left)) but the VPdl
neuron (Figure 7B (left)) exhibited an increase in FR just prior to the approach offset. Analysis of the directional changes in FR during the response revealed that the VPdl was significantly decreased in FR compared to the VPvm, \( z = -2.18, p < 0.05 \). The VPdl median directional change in FR during the response was 0.44 (approximately a 21% reduction in FR) and median VPvm directional change in FR was 0.49 (less than 1% reduction in FR).

To examine the influence of the S\(^D\) upon response-related firing patterns, responses were sorted into cued and uncued trials. Between subregions, the VPdl exhibited larger cued (Figure 7D) and uncued (Figure 7E) absolute changes in FR than the VPvm (cued \( z = -2.68, p < 0.01 \); uncued \( z = -2.31, p < 0.05 \)).

Within the VPdl alone, a significant correlation was observed between cued and uncued response directional changes in FR, VPdl \( r = 0.38, p < 0.01 \) (Figure 7F).

Within the VPvm alone, cued and uncued response directional changes in FR were also significantly correlated, VPvm \( r = 0.56, p < 10^{-14} \) (Figure 7G). The correlation coefficients did not differ between subregions, \( z = -1.33, p > 0.05 \).

That is, within each subregion neurons did not discriminate changes in FR between cued response and uncued response conditions (examples are displayed in Figure 7H for a VPdl neuron and Figure 7I for a VPvm neuron).

Outliers within Figures 7F-G suggested a very small number of neurons discriminated between cued and uncued response conditions. However, visual inspection of these neurons’ changes in FR determined that differences between cued and uncued response conditions were due to directional change comparisons \( (B/(A+B)) \) which exhibited low FRs in both 'A' and 'B' conditions.
(i.e., FR < 0.1 discharges/sec that otherwise met all criteria for inclusion as individual units) that differed only slightly, rather than exhibiting clear patterned changes in FR during only cued or only uncued response. Thus, similar to the approach, during cocaine self-administration the SD has little to no influence upon response-related firing patterns within neurons from either VP subregion.

No VP neurons exhibited any significant relationship of FR with vertical head movement properties including distance (Figure 8A), duration (Figure 8B), velocity (Figure 8C), start position/valley (Figure 8D), or end position/peak (Figure 8E). This is in contrast to head-movement neurons of the dorsolateral striatum, which exhibit exquisite sensitivity to these movement parameters with FR, as measured during similar motivated behavior (Pederson et al. 1997; Tang et al. 2007; Pawlak et al. 2010).

Fewer neurons exhibited their only change in FR during the retreat away from the photocell corner (Table 4). No significant differences were found between subregions for the absolute changes in FRs during the retreat (z = -0.92, p > 0.05; Figure 9A). Of neurons that exhibited a retreat-only firing pattern, these neurons were typically located in the VPvm (Table 4; example neuron shown in Figure 9B). In an attempt to identify the influence of cocaine infused prior to or during the retreat on changes in FR during retreat behavior, retreats were sorted into trials in which the pump was activated two seconds prior to the onset of retreat through the offset of the retreat versus all other retreats, termed pump retreats and nonpump retreats, respectively.
Within VPdl alone, a significant correlation was observed between pump and nonpump retreat directional changes in FR, VPdl $r = 0.59$, $p > 10^{-4}$ (Figure 9C). Within the VPvm alone, pump and nonpump retreat directional changes in FR were also significantly correlated, $r = 0.40$, $p < 10^{-6}$ (Figure 9D). The correlation coefficients did not differ between subregions, $z = 1.65$, $p > 0.05$. That is, VP neurons’ changes in FR largely did not discriminate between retreats with or retreats without pump activation. Two examples of similar changes in FR between pump and nonpump retreat conditions are shown for a VPdl neuron in Figure 9E and VPvm in Figure 9F. Nevertheless, some neurons exhibited differences in FRs between pump and nonpump retreat conditions in both subregions (VPdl neuron example in Figure 9G and VPvm neuron example in Figure 9H). It is possible that this minority of neurons plays a role in reward prediction, or may be related to other factors not investigated here. In general, there appears to be little influence of the initial seconds of cocaine infusion prior to or during the retreat on changes in FR during retreat behavior.

In summary, 37.50% of VPdl and 39.77% of VPvm neurons exhibited at least a 20% change in FR during only the approach, only the response, or only the retreat (Table 4), suggesting that the behaviorally-coincident firing patterns observed in NAcc neurons during cocaine self-administration (Chang et al. 1994, 1997, 2000) are projected to VP. VPdl neurons exhibited a significantly larger change in FR during approach or response than VPvm neurons, whether such approaches or responses were cued or uncued. Given that the NAcc core projects to the VPdl and NAcc shell projects to the VPvm, these results extend
the repeated observation that core neurons exhibit greater changes in FR during cocaine-seeking behaviors than shell neurons (Ghitza et al. 2004; Hollander and Carelli, 2005; Fabbricatore et al. 2010). No differences between subregions were observed for changes in FR during the retreat. VP neurons did not discriminate between cued and uncued conditions for the approach or response and largely did not discriminate between retreats involving pump infusions or not. The $S^D$ has little behavioral relevance during cocaine self-administration (Root et al. 2011), in contrast to $S^D$ effects on abstinent drug-seeking (Ghitza et al. 2003; Root et al. 2009). The fact that VP changes in FR during approach and response during self-administration do not differ as a function of cue presentation suggests that VP neurons are engaged during conditions that are behaviorally-relevant rather than those that are not relevant.

Changes in FR during two of three cocaine-seeking behaviors

We next examined whether VP neurons discriminated two of the three cocaine-seeking behaviors. Neurons that exhibit changes in FR during both approach and response might be related to processing the execution of drug-seeking and taking behaviors. Neurons that exhibit changes in FR during both approach and retreat, or during both response and retreat, might be involved in the association of cocaine-seeking behaviors with their expected reinforcement by cocaine.

When a change in FR occurred during the approach, VPdl neurons were highly likely to continue this change in FR in the same direction with similar or...
greater magnitude through the duration of the response (described briefly above for the VPdl neuron in Figure 7B). Within the VPdl, a significant correlation was observed between the directional changes in FR during the approach and response, $r = 0.69$, $p < 10^{-7}$ (Figure 10A). Two examples of VPdl neuron approach-response firing patterns are shown in Figures 10B-C. While a significant correlation between the directional changes in FR during the approach and response was also observed within the VPvm, $r = 0.32$, $p < 10^{-4}$ (Figure 10D), VPdl exhibited a significantly larger correlation coefficient than VPvm, $z = 3.11$, $p < 10^{-3}$. Using the response directional change in FR as a dependent variable, this result was corroborated by an observed interaction between approach directional change in FR and subregion, $F(1, 215) = 10.81$, $p < 0.01$.

That is, the linear slopes of VPdl and VPvm neurons' relationship between approach and response differed between subregions. Taken together, although some VPvm neurons continued to change their FRs in the same direction from approach through response (example VPvm neuron in Figure 10E), the weaker correlation of approach and response directional changes in FR by VPvm neurons suggests this subregion exhibits more behavioral selectivity and thus more heterogeneous firing patterns, including approach-only or response-only changes in FR. In contrast, the stronger correlation of approach and response directional changes in FR by VPdl neurons suggests this subregion may not typically discriminate drug-seeking from drug-taking during self-administration.

For neurons that exhibited a change in FR during the retreat, these were largely coupled with changes in FR during either the approach, response, or both
the approach and response (Table 4). A significant correlation was observed for the directional changes in FRs during the approach and retreat within the VPdl alone, \( r = 0.51, p < 0.001 \) (Figure 11A) and within the VPvm alone, \( r = 0.41, p < 10^{-7} \) (Figure 11B). Subpopulations of VP neurons exhibited a change in FR during approach, reverted back to baseline during the response, and exhibited another change in FR during the retreat (Figures 11C-D; Table 4).

Other subpopulations of VP neurons exhibited changes in FR during response and retreat (Table 4). A significant correlation was observed for the directional changes in FRs during the response and retreat within the VPdl alone, \( r = 0.37, p < 0.01 \) (Figure 11E) and within the VPvm alone, \( r = 0.19, p < 0.05 \) (Figure 11F). Two examples of response-retreat firing patterns are displayed in Figure 11G for a VPdl neuron and Figure 11H for a VPvm neuron. The correlation coefficients for approach-retreat and response-retreat did not differ between subregions (all \( |z| < 0.74, p > 0.05 \)).

In summary, 25.00% of VPdl and 26.32% of VPvm neurons exhibited at least a 20% change in FR involving two behaviors, approach and response, approach and retreat, or response and retreat. VPdl neurons exhibited a significantly larger correlation coefficient between directional changes in FR between approach and response than VPvm neurons, suggesting VPdl neurons uniquely participate in the execution of drug-seeking behaviors. The correlation coefficients of approach and retreat as well as response and retreat did not differ between subregions.
Changes in FR during all three cocaine-seeking behaviors

We next examined whether VP neurons exhibited changes in FR during all three behaviors. Such changes in FR might be considered a "gating" signal if they occur in the same direction (all increases or all decreases in FR), which has been observed in NAcc neurons (Taha and Fields, 2006). In contrast, such changes in FR might signal the contrasting components of different cocaine-seeking behaviors if they occur in heterogeneous directions.

Approximately 22.92% of VPdl and 9.94% of VPvm neurons exhibited at least a 20% change in FR during all three behaviors. Most of these neurons changed FR beginning at the onset of approach and continued in the same direction (increase or decrease in FR) through the response and retreat (Table 4; VPdl neuron example in Figure 12A and VPvm neuron example in Figure 12B), suggesting the possibility of a "gating" signal. A smaller population of neurons that exhibited a change in FR during approach, response, and retreat did so with heterogeneous directional changes in FR (Table 4; Figures 12C-D show VPdl neurons and Figures 12E-H show VPvm neurons).

Cue

Clear examples of changes in FR following SD onset were not observed. With respect to absolute changes in FR in response to the cue, no significant differences were found between subregions (z = -0.93, p > 0.05) (data not shown). After sorting cue trials into those in which the animal self-administered cocaine (hits) versus those in which the animal did not self-administer cocaine
Ventral pallidum subregions

(misses), no differences between subregions were found for the absolute changes in FR (all $|z| < 1.16$, $p > 0.05$). Furthermore, no clear examples of changes in FR during hit or miss trials were observed. In summary, VP neurons were not sensitive to the onset of the $S^D$ during cocaine self-administration, whether the trial was reinforced or not.

Examination of “initiation”-related firing patterns

The final analysis investigated whether VP neurons exhibited firing patterns prior to approach onset (i.e., pre-movement firing). Such changes in FR have been observed in NAcc neurons (Chang et al. 1994) but were tacitly postulated to occur in VP neurons during the "initiation" of motivated behaviors (Mogenson et al. 1980). Visual inspection of all neurons’ firing patterns revealed only three candidate neurons with changes in FR that began prior to the approach onset (Figure 13A (left), 14B (left), 14C (left)). The two neurons displayed in Figures 13A-B were recorded from the same rat and we returned to the recorded videos to reanalyze the self-administration behaviors. This rat exhibited a locomotor movement (alternating limb movements) toward a specific corner of the chamber just prior to the approach toward the photocell corner, which was termed the pre-approach movement (Movie 2). Overlaying the pre-approach movement onset and offset (magenta and cyan dots in Figures 13A (right) and 14B (right), respectively) over the two candidate neuron rasters clearly demonstrated that the decrease (Figure 13A (second from right)) or increase (Figure 13B (second from right)) in FR prior to approach (time zero) was related
to the pre-approach movement. The final candidate neuron, shown in Figure 13C, was recorded in a different rat and exhibited a decrease in FR prior to the approach onset (Figure 13C (left)). For this neuron, we examined whether previous approaches (Figure 13C (third from right)), responses (Figure 13C (second from right)), or retreats (Figure 13C (right)) explained the decrease in FR prior to the approach. Figure 13C (right) demonstrates that this rat often reapproached the operandum quickly following retreat offset, and that a decrease in FR occurred in these trials near the offset of retreat. In summary, with respect to the behaviors studied in the present cocaine self-administration task, changes in FR of 219 VP neurons in no case preceded the onset of approach but rather occurred strictly during approach, response, and/or retreat behaviors.

DISCUSSION

A major goal in systems neuroscience and neurobiology of drug abuse is the characterization of the unique contributions of subregionally specific brain regions that underlie drug-seeking behavior. The present results extend neuroanatomical studies of VP subregional afferent and efferent projection patterns (Zahm and Heimer, 1988, 1990; Groenewegen et al. 1993; Bell et al. 1995; Kalivas et al. 1993; Zahm et al. 1996; Churchill et al. 1996; Heimer et al. 1991, 1997; O'Donnell et al. 1997; Tripathi et al. 2010) by demonstrating differential changes in FR within VPdl and VPvm subregions during distinct components of cocaine-seeking behavior. Neurons within the calbindin-d28k immunoreactive VPdl exhibited a greater absolute change in FR during the
approach and response relative to neurons in the neurotensin-immunoreactive VPvm. This was the case for cued and uncued approaches as well as cued and uncued responses. Since the NAcc core projects to the VPdl (Zahm and Heimer, 1990; Groenewegen et al. 1993; Zahm et al. 1996; O'Donnell et al. 1997; Tripathi et al. 2010), and core neurons exhibit a greater change in FR over medial NAcc shell neurons during cocaine-seeking behavior (Ghitza et al. 2004, 2006; Hollander and Carelli, 2005; Fabbricatore et al. 2010), the NAcc core - VPdl subcircuit may be an especially important contributor to the approach and response components of drug-seeking behavior.

The patterns in which changes in FR occurred also differed by subregion. The increases/decreases in FR during approach and response of VPdl neurons exhibited a significantly larger correlation coefficient than VPvm neurons, indicating that VPdl neurons were more rigid in their approach and response firing patterns. That is, changes in FR by VPdl neurons typically began during the approach and carried through the response in the same direction with similar or greater magnitude. In contrast, VPvm neurons were heterogeneous, changing FRs during the approach alone, response alone, or both approach and response. Taken together with the greater magnitude change in FR during approach and response by VPdl neurons compared to VPvm neurons, distinct firing patterns during these behaviors suggests VP subregions play differential roles during drug-seeking behavior.

The VPdl, with weak projections to MDT and robust efferents to "motoric" regions such as subthalamic nucleus and medial substantia nigra pars reticulata
Ventral pallidum subregions may be an important contributor to the expression of drug-seeking and drug-taking behaviors via its approach and response firing patterns. Fan et al. (2012) recently reported that many medial substantia nigra neurons (especially putative GABAergic neurons) exhibit changes in FR during responses. Our results suggest the VPdl is one source of afferent input to substantia nigra pars reticulata neurons regulating changes in FR during the response. Furthermore, given that NAcc core neurons exhibited predominant increases in FR over NAcc shell neurons during cocaine-seeking (Ghitza et al. 2004) and VPdl neurons were significantly decreased in FR compared to VPvm neurons during the response, the influence of VPdl GABAergic efferents upon their targets during the response may be through disinhibition.

The VPvm exhibited heterogeneous firing patterns, including approach alone, response alone, and approach and response. The capacity of VPvm neurons to differentiate approach from response (ie, drug-seeking from drug-taking) is consistent with greater convergent limbic signaling in the medial corticostriatopallidal circuit (French and Totterdell, 2002, 2003, 2004). Taken together with its robust innervation of the MDT and VTA (Zahm and Heimer, 1990; Groenewegen et al. 1993; Zahm et al. 1996; O'Donnell et al. 1997), heterogeneous firing patterns may implicate VPvm neurons in facilitating mesocortical structures with information related to different temporal components or sequences of behaviors that predict reward. In support, VP, MDT, and medial prefrontal cortex lesions disrupt performance following behavioral contingency...
changes (i.e. reversal learning or switching from Pavlovian to operant contingencies) where monitoring of new appropriate behavioral sequences (approach, response, retreat) are critical (McBride and Slotnick 1997; Ferry et al. 2000; Block et al. 2007; Pickens, 2008). Other firing patterns, also heterogeneous in direction, were observed in the VPvm more than the VPdl, such as retreat alone as well as response-retreat. Such firing patterns may be involved the association of cocaine-taking behaviors and their anticipated reinforcement. The NAcc shell, but not core, supports intracranial self-administration of a variety of drugs of abuse (Carlezon et al. 1995; Carlezon and Wise, 1996a,b; Ikemoto et al. 1997, 2005; Rodd-Henricks et al. 2002; Shin et al. 2008), suggesting the VPvm is involved in reward-related mechanisms.

In addition to heterogeneous firing patterns of shell neurons during cocaine self-administration (Ghitza et al. 2004; Fabbricatore et al. 2010) other factors may account for the heterogeneous directional changes in FR of VPvm neurons, such as the differential mean area densities of dopamine transporter- and tyrosine hydroxylase-labeled axons between VP subregions (Mengual and Pickel, 2004) or collaterals into VPvm from VPdl-projecting core neurons (Zahm and Brog, 1992; Brog et al., 1993; Tripathi et al. 2010), Additional sources of heterogeneity, which would affect both VP regions, may result from chronic cocaine exposure which alters GABA, opioid, and glutamate-induced changes in FR of VP neurons (McDaid et al., 2005); excitatory substance P projections from NAcc (Mitrovic and Napier, 1998) which rarely colocalize with GABAergic or enkephalinergic in VP terminals (Zahm et al., 1985); and elevated extracellular
dopamine or serotonin concentrations in VP during cocaine self-administration (Sizemore et al. 2000) which are robust modulators of VP FRs (Napier et al., 1991; Napier and Maslowski-Cobuzzi, 1994; Heidenreich and Napier, 2000).

Whether or not the above manipulations differentially affect VP subregions is untested.

In primarily the VPvm, a small population of neurons exhibited changes in FR only during the retreat away from the self-administration operandum. The absolute change in FR during the retreat did not differ between subregions. The retreat-related changes in FR observed here extend the "receptacle exit" firing patterns of NAcc neurons during natural reward-seeking behavior (Nicola et al. 2004a,b; Yun et al. 2004). It is likely that some retreat-related changes in FR reflect the "post-press" changes in FR (e.g. following response completion) observed in our previous examination of VP neurons using a lever press operant (Root et al. 2010). However, some firing patterns occurred after the retreat offset (Figures 7B (right), 10C (right)). Within each subregion, the directional change in FR during retreats involving pump onset significantly predicted the directional change in FR during retreats that did not contain pump onset, suggesting on the whole that the cocaine infusion did not induce a rapid phasic change in FR. The behaviorally-relevant timescale for which our analyses occurred, hundreds of milliseconds (Table 3), occur too rapidly for pharmacologically-induced changes in FR (discussed in Root et al. 2010). We have recognized pharmacological changes in FR by VP neurons in a previous report (Root et al. 2012), but these changes in FR occur over significantly longer timescales (minutes to hours). For
the minority of neurons that exhibited differential firing patterns during pump versus nonpump retreats, it is unclear whether retreats differ behaviorally between pump and nonpump conditions. Other factors, such as visceral effects of pump activation or potential success or error signals may contribute to firing patterns during the pump or nonpump retreat conditions. Furthermore, the retreat might reflect the anticipation of forthcoming cocaine reward and/or a behavior that leads to the onset of focused stereotypy. Further experimentation will be necessary to detail the factors involved in retreat-related firing patterns.

We observed no changes in FR related to $S^D$ presentations. NAcc neurons exhibit a paucity of $S^D$-related changes in FR during cocaine self-administration (unpublished observations). One reason for the lack of changes in FR induced by the $S^D$ may be that the $S^D$ has little measurable behavioral influence while the animal is self-administering cocaine. During cocaine self-administration, responses occur only when the drug level falls below a unique individual internal threshold, above which the animal is satiated and does not respond (Pickens and Thompson 1968; Wise 1987; Lynch and Carroll 2001; Norman and Tsibulsky 2006). We have found evidence of such behavior using the same head movement paradigm employed in the present study. Specifically, clamping each animal's drug level immediately above its satiety threshold eliminated responding while clamping drug level below satiety threshold markedly increased responding above baseline, irrespective of $S^D$ presentations in both tests (Root et al. 2011). In other words, during cocaine self-administration the animal's current drug level overshadows the influence of the $S^D$ upon behavior. Consistent with the inability
of the $S^D$ to guide behavior during self-administration, it is then not surprising that VP neurons did not discriminate cued approaches from uncued approaches as well as cued responses from uncued responses. We suggest that changes in FR during approach and response are nevertheless “cue-induced” by the crossing of cocaine level below an internal threshold (Root et al., 2012), but not by the external $S^D$ during self-administration. Whether or not VP neurons discriminate cued from uncued behaviors during natural reward seeking behavior, akin to NAcc neurons (Nicola et al. 2004a), is untested.

While we and others observe no behavioral evidence that the $S^D$ influences behavior during self-administration, we have found that the animals nevertheless learn the relationship of $S^D$ with the outcome of cocaine infusions. When the animal is abstinent, responding is low when the $S^D$ is off and significantly increased following $S^D$ presentation (Ghitza et al. 2003; Root et al. 2009). Furthermore, at this time when the $S^D$ is behaviorally relevant (abstinent testing), NAcc neurons exhibit robust changes in FR following $S^D$ presentation (Ghitza et al. 2003). Whether or not VP neurons show $S^D$-induced changes in FR during abstinence remains to be investigated.

The ventral striatopallidal system is hypothesized to be involved in goal-directed behaviors (Mogenson et al. 1980; Alexander et al. 1986). During cocaine self-administration, the same NAcc neurons that changed FR during approach and response did not exhibit changes in FR during locomotion induced by a treadmill (Chang et al. 1994), demonstrating “goal-directed” rather than “locomotor” firing patterns. While the present study did not utilize a treadmill, we
argue that the changes in FR observed in VP neurons were also “goal-directed”. Given that i) 66.67% of VPdl and 53.22% of VPvm neuron firing patterns included the response, ii) the response in the present vertical head movement operant task precluded locomotion, and iii) no VP neurons exhibited significant correlations between FR and any motoric parameter of the response (e.g., distance, duration, velocity, valley, apex), other factors must be involved in producing the observed VP firing patterns during approach, response, and retreat. Furthermore, the lack of strictly “motoric” or “locomotor” firing correlates by VP neurons extends similar results from other laboratories. Mitchell and colleagues (1987) reported VP neurons i) exhibit no changes in FR in response to passive or active movement of any body part prior to conditioning and ii) do not encode information regarding direction and muscle pattern of movements following conditioning. Tindell and colleagues (2004) reported VP neurons do not exhibit changes in FR during “right and left stepping movements”, i.e. locomotion.

The NAcc is the primary afferent of the VP (Zahm et al. 1985). Presumably, the “goal-directed” firing patterns observed in NAcc (Chang et al. 1994) are projected to the VP. Based on their groundbreaking studies of changes in FR during specific body part movements that occurred in serial fashion across interconnected regions of the “skeletomotor” dorsolateral striatal system, Alexander and colleagues (1986) predicted that firing patterns were similarly serially transmitted within the "cingulate" system; consisting of cortical projections to NAcc, followed by NAcc projections to VP, and subsequent VP projections to MDT. Seminal studies by Woodward and colleagues, recording medial prefrontal
cortex (Chang et al. 1997, 2000) and NAcc (Chang et al. 1994, 2000) neurons during cocaine self-administration, together with the current VP study, provide critical insight into the function of this circuit. In their studies, the lever was positioned such that in order to respond, rats had to approach the lever, rear, and position their forelimbs to press the lever downward. Firing patterns observed in their recordings of mPFC and NAcc were exclusively related to approaching the lever alone or to aspects involving the approach to the lever (rearing) and responding (touching the lever). Given that the most common firing patterns observed within the VP were related to approaching or responding alone on the operandum as well as approaching and responding together, the present results support the hypothesis (Alexander et al. 1986) of a maintained afferent firing pattern involving approach and response within the cingulate circuit. Taking the above arguments together with the similar firing patterns observed between the present experiment utilizing a vertical head movement operant and our prior investigation of VP firing patterns using a different motoric requirement for cocaine self-infusions (lever press operant; Root et al. 2010), VP neurons appear to acquire firing patterns with one or more learned goal-directed behaviors involved in self-administering cocaine, regardless of motoric detail per se.

Whether or not VP passively relays signals from NAcc is not clear. The NAcc projects GABA to VP (Haber and Nauta 1983; Zahm et al., 1985). However, the projection is predominantly opioid colocalized (Zahm et al., 1985; Olive et al., 1997), which is antagonistic to the effects of GABA (Chrobak and Napier, 1993). Other NAcc projections to VP contain substance P (Haber and
Nauta, 1983; Zahm et al. 1985; Napier et al., 1995), which are excitatory on VP neurons (Napier et al., 1995; Mitrovic and Napier, 1998). With respect to cocaine-seeking behaviors, it appears that the VP (especially the VPvm) is more directionally heterogeneous in its firing patterns than NAcc neurons. We can only speculate that the signals from NAcc reach VP with additional heterogeneity due to colocalization and influences from other afferents. Our data do not resolve the mechanisms with which signals are processed within ventral striatopallidal pathways and future research will be necessary.

Differences were observed between the present experiment and those reported by Tindell and colleagues (2004). The lack of cue-induced changes in FR observed in the present experiment but reported by Tindell et al. (2004) during a Pavlovian sucrose task was likely due to the cocaine self-administration paradigm (explained above). However, Tindell and colleagues (2004) also reported that only 3.03% (3/66) of VP neurons that exhibited CS+ or CS- related activity, also exhibited a change in FR related to approaching the reward receptacle. Several reasons may explain this discrepancy. First, in contrast to Tindell et al. (2004), the present experiment utilized immunohistochemistry as well as organized arrays of microwires rather than bundled electrodes. The present methods unequivocally localized every microwire recording reported in the present experiment to within the VP. While VP is a predominantly subcommissural structure (Heimer and Wilson, 1975; Heimer, 1978; Heimer et al. 1997), the recordings of the Berridge/Aldridge labs (Tindell et al. 2004, 2005, 2006; Smith et al. 2011) are located near or within the caudal, sublenticular parts
of VP. Zahm and colleagues (2012) recently noted that the sublenticular regions "said to be located in the posterior part of the ventral pallidum may actually significantly involve the lateral preoptic area or a transitional merging of the ventral pallidum and lateral preoptic area." Thus, histological location of recorded neurons may be one reason for differences between the present study and those by the Berridge/Aldridge labs. Second, NAcc neurons, the primary afferent of VP neurons, exhibit changes in FR during approach/response or "pre-response" epochs using cued (S^D or Pavlovian) as well as uncued natural reward paradigms (Carelli and Deadwyler, 1994; Carelli et al. 2000; Chang et al. 2002; Nicola et al. 2004a,b; Yun et al. 2004; Wood et al. 2004; Day et al. 2006; Jones et al. 2010). While this suggests VP neurons would exhibit approach/response-related firing patterns, as we verify in the present report during cocaine self-administration, it is plausible that differential firing patterns occur within VP neurons depending on the interaction of reward contingency (Pavlovian or Skinnerian) and type of reward (natural or drug reward).

Finally, of critical importance for drug-abuse research is the determination of which brain regions are responsible for initiating and executing drug-seeking behaviors. Mogenson and colleagues (1980) theorized that through projections to the pedunculopontine tegmental mesencephalic locomotor region, VP provided a gateway for limbic/emotional signals sent to NAcc to gain access to the motor system. If VP signals the "initiation" (Mogenson et al. 1980) of motivated behavior, one would expect firing pattern changes prior to the onset of approach. Only 3 candidate VP neurons exhibited changes in FR prior to the approach,
which upon further inspection were revealed to be due to other factors than an "initiation" firing correlate. Instead of preceding the onset of approach, VP FR changes in every case were coincident with approach, response, and/or retreat behaviors. Taken together with the arguments of Heimer and colleagues (1997), who contended that previous investigations of VP projections to the mesencephalic locomotor region "lumped" the VP with the subpallidal parts of the extended amygdala, VP is not likely the "initiator" of cocaine-seeking behavior.

Instead, the role of VP upon cocaine self-administration behaviors is subregionally dependent. The stronger, sustained FR changes of VPdl neurons during approach and response may implicate VPdl in similarly processing the acts of drug-seeking and drug-taking behaviors via projections to subthalamic nucleus and substantia nigra pars reticulata. In contrast, heterogeneous firing patterns of VPvm neurons may provide mesocortical structures with information related to the sequence of behaviors predicting cocaine self-infusions via projections to MDT and VTA. Thus, the present results together with the neuroanatomical literature provide a neurophysiological basis for functionally distinct but integrative involvement of VPdl and VPvm subregions in addiction.
References


expression in a mouse model of mesial temporal lobe epilepsy.

Hippocampus 10:305-324.


Geisler S, Zahm DS. 2006b. On the retention of neurotensin in the ventral tegmental area (VTA) despite destruction of the main neurotensinergic afferents of the VTA—Implications for the organization of forebrain projections to the VTA. Brain Res 1087:87-104


Ikemoto, Glazier BS, Murphy JM, McBride WJ. 1997. Role of dopamine D1 and D2 receptors in the nucleus accumbens in mediating reward. JNeurosci 17(21):8580-8587


Morse DT. 1999. MINSIZE2: a computer program for determining effect size and minimum sample size for statistical significance for univariate, multivariate, and nonparametric tests. Educ Psychol Meas 59: 518–531

Napier TC, Maslowski-Cobuzzi RJ. 1994. Electrophysiological verification of the presence of D1 and D2 dopamine receptors within the ventral pallidum. Synapse 17:160–166.


Nicola SM, Yun IA, Wakabayashi KT, Field HL. 2004a. Firing of nucleus accumbens neurons during the consummatory phase of a discriminative stimulus task depends on previous reward predictive cues. J Neurophysiol 91: 1866:1882


Peoples LL, Uzwiak AJ, Gee F, West MO. 1997. Operant behavior during sessions of intravenous cocaine infusion is necessary and sufficient for


Ventral pallidum subregions

accumbens shell, core, and olfactory tubercle. Psychopharmacology 198(2):261-270


Zahm DS, Zaborszky L, Alones VE, Heimer L. 1985. Evidence for the coexistence of glutamate decarboxylase and met-enkephalin


Zahm DS, Parsely KP, Schwartz ZM, Cheng AY. (2012). On lateral septum-like characteristics of outpute from the accumbal hedonic 'hotspot' of Pecina
and Berridge with commentary on the transitional nature of basal forebrain ‘boundaries’. J Comp Neurol, in press.
TABLE LEGENDS

Table 1. For further details regarding the primary antibodies, see the Antibody Characterization section.

Table 2. Units and Triphasic refer to total number and percent, respectively. All other data are presented as means ± SEM. *Amplitude of unitary action potentials. VPvm- ventromedial ventral pallidum; VPdl- dorsolateral ventral pallidum.

Table 3. Data are presented as mean ± SEM number of events per animal as well as mean ± SEM duration (ms).

Table 4. Data are presented as percent and total number of neurons, respectively, within a particular VP subregion (VPdl or VPvm). Totals refer to number of neurons exhibiting 20% change from baseline. This categorization was intended for simplification purposes that were sensitive enough to detect relatively small changes in FR.
FIGURE LEGENDS

Figure 1. Behaviors schematized for neuronal analysis. Appetitive behaviors included the approach towards the photocell corner (A), criterion head movement operant response (B), and retreat away from the photocell corner (C). Arrows in (A) and (C) indicate locomotion that is absent from (B). The criterion movement consisted of a vertical head movement breaking photocells 1 or 2 through 5 or 6, consecutively, under one second in duration. In this example, the rat executes a vertical head movement breaking photocells 1 through 6.

Figure 2. Self-administration and cue learning. Reaction time in response to the $S^D$ (A), probability of self-administering cocaine upon $S^D$ presentation (B), cocaine consumption (C), latency to load up to the tenth infusion (D), number of criterion movements (E). Values are average ± SEM (y axes) per day (x axes).

Figure 3. Skill learning: changes in criterion head movement parameters with extensive repetition. Changes in velocity (A), duration (B), distance (C), probability of starting at photocell two, the minimum required for a criterion movement (D), and the probability of ending at photocell 5 (E). Values for A, B, and C are average median ± SEM per day for criterion head movements (y axes). Values for D and E are average probability ± SEM. All x axes refer to training day.
Figure 4. Example microwires localized to VP subregions. Examples are displayed in groups of three. A-I displays three examples of microwires localized to the VPdl in three vertical panels each; example one A-C, example two D-F, and example three G-I. J-R display three examples of microwires localized to the VPvm in three vertical panels each; example one J-L, example two M-O, and example three P-R. Panels A, D, G, J, M, and P display substance P immunohistochemistry. Panels B, E, H, K, N, and Q display calbindin-d28k immunohistochemistry. Panels C, F, I, L, O, and R display neurotensin immunohistochemistry. Green/blue dot in each panel is an iron deposit from the uninsulated microwire tip visualized from by potassium ferrocyanide counterstain.

Numbers refer to approximate anteroposterior coordinate based on Paxinos and Watson (2004). As a model for all VPdl neurons recorded, VPdl example 1 (A-C) displays two microwire tip locations. The wire closest to the anterior commissure was localized to substance P immunoreactivity (A) and calbindin-d28k immunoreactivity (B), but not neurotensin immunoreactivity (C). The microwire furthest from the anterior commissure (A) was localized outside of substance P immunoreactivity and therefore excluded from the dataset. As a model for all VPvm neurons recorded, VPvm example 1 (J-L) displays a microwire tip location localized to substance P immunoreactivity (J) and neurotensin immunoreactivity (K), but not calbindin-d28k immunoreactivity (L). Any tissue from the left hemisphere has been rotated horizontally to the right hemisphere for the sake of uniform orientation (midline is left for all panels). Outlines were based on substance P, calbindin-d28k, and neurotensin immunoreactivity, the atlas of
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Calibration bar in A represents 1 mm. All sections were 40 µm thick.

Figure 5. All microwires from all rats that were localized to VP subregions. Tissue displayed is substance P immunoreactivity from the left hemisphere of a rat used in the present dataset. The left hemisphere implant from this rat was misplaced dorsal to VP and thus unencumbered by microwire tip counterstaining, and therefore was selected for display purposes. Black circles and cyan triangles indicate VPvm and VPdl localized microwires, respectively. A-J display right hemisphere localized microwires (midline right). K-U display left hemisphere localized microwires (midline left). Outlines were based on substance P immunoreactivity as well as the atlas of Paxinos and Watson (2004). Numbers refer to approximate anteroposterior coordinate based on Paxinos and Watson (2004). Calibration bar in A represents 1 mm. All sections were 40 µm thick.

Figure 6. VPdl neurons exhibit greater changes in FR during the approach than VPvm neurons whether cued or uncued. (A) Cumulative proportion of all neurons' absolute changes in FR during the approach per subregion. Gray and black lines indicate cumulative proportions for VPdl and VPvm neurons, respectively. Median values (0.50 proportion value) are displayed as circle (VPdl) and square (VPvm). For this and all other cumulative proportion figures, the absolute changes in FR were used. A value of 0 was no difference from baseline FR and a
fifty percent change from baseline FR was 0.1. Examples of approach-related changes in FR are displayed in (B) from a VPdl neuron and (C) from a VPvm neuron. For (B) and (C), rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots in approach-raster and blue dots in response-raster indicate the onset of approach and onset of response, respectively. Red dots in retreat-raster indicate the offset of retreat. The cumulative proportions of all neurons' absolute changes in FR for cued approach and uncued approach per subregion are shown in (D) and (E), respectively. Each VPdl (F) and VPvm neuron (G) directional change in FR during the cued approach (y-axis) was plotted against its directional change in FR during the uncued approach (x-axis). Each dot represents one neuron. Note that for (F) and (G), and all other scatter plot figures, directional changes in FR were used. In contrast to the absolute change in FR, a value of 0.5 was not different from baseline FR and a fifty percent change from baseline FR was 0.6 (increase) or 0.4 (decrease). Two examples of similar directional changes in FR between cued and uncued approach changes in FR, one VPdl neuron (H) and one VPvm neuron (I) are displayed. For (H) and (I), rasters and PETHs are centered (time 0) around offset of cued approach (left) and uncued approach (right). Green dots indicate the onset of approach. All rasters have been sorted for movement duration. Y-axes of all rasters refer to behavioral trial; y-axes of all PETHs refer to average FR (spikes/sec) per trial. All raster and PETH x-axes refer to time (ms). *** p < 0.001, ** p < 0.01; * p < 0.05.
Figure 7. VPdl neurons exhibit greater changes in FR during the response than VPvm neurons whether cued or uncued. (A) Cumulative proportion of all neurons' absolute changes in FR during the response per subregion. Gray and black lines indicate cumulative proportions for VPdl and VPvm neurons, respectively. Median values (0.50 proportion value) are displayed as circle (VPdl) and square (VPvm). Examples of approach-related changes in FR are displayed in (B) from a VPdl neuron and (C) from a VPvm neuron. For (B) and (C), rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots in approach-raster and blue dots in response-raster indicate the onset of approach and onset of response, respectively. Red dots in retreat-raster indicate the offset of retreat. The cumulative proportion of all neurons' absolute changes in FR for cued approach and uncued approach per subregion are shown in (D) and (E), respectively. Each VPdl (F) and VPvm neuron (G) directional change in FR during the cued approach (y-axis) was plotted against its directional change in FR during the uncued approach (x-axis). Each dot represents one neuron. Two examples of similar directional changes in FR between cued and uncued approach changes in FR, one VPdl neuron (H) and one VPvm neuron (I) are displayed. For (H) and (I), rasters and PETHs are centered (time 0) around offset of cued response (left) and uncued response (right). Blue dots indicate the onset of response. All rasters have been sorted for movement duration. Y-axes of all rasters refer to behavioral trial; y-axes of all PETHs refer to average FR (spikes/sec) per trial. All raster and PETH x-axes refer to time (ms). ** p < 0.01; * p < 0.05.
Figure 8. No relationship between response movement parameters and changes in FR by VP neurons. Given that no significant relationships were observed, all neurons were pooled into graphs of distance (A), duration (B), velocity (C), valley (start position) (D), and peak (end position) (E) regardless of subregion. Each bin on the x-axis is 0.10. Y-axis is frequency of occurrence (number of neurons exhibiting the r value).

Figure 9. No differences between subregions during the retreat. Cumulative proportion of all neurons’ absolute change in FR during the retreat per subregion. Gray and black lines in (A) indicate cumulative proportions for VPdL and VPvm neurons, respectively. Median values (0.50 proportion value) are displayed as circle (VPdL) and square (VPvm) in (A). One example of retreat-related firing is displayed in (B), from a VPvm neuron. Raster and PETH are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dot in approach:raster and blue dot in response:raster indicate the onset of approach and response, respectively. Red dot in C indicates the offset of retreat. For each VPdL (C) and VPvm neuron (D), the neuron’s directional change in FR during the nonpump retreat (y-axis) was plotted against its directional change in FR during the pump retreat (x-axis). Each dot represents one neuron. Two examples of similar directional changes in FR between pump retreat and nonpump retreat changes in FR, one VPdL neuron (E) and one VPvm neuron (F) are displayed. Two examples of dissimilar directional changes in FR
between pump retreat and nonpump retreat changes in FR, one VPdl neuron (G) and one VPvm neuron (H) are displayed. For (E), (F), (G), and (H), rasters and PETHs are centered (time 0) around onset of pump retreats (left) and nonpump retreats (right). Red dots indicate the offset of the retreat. All rasters have been sorted for movement duration. Y-axes of rasters refer to behavioral trial; y-axes of PETHs refer to average FR (spikes/sec) per trial. Both x-axes refer to time (ms).

Figure 10. VPdl neurons exhibit a greater correlation between approach-and-response changes in FR than VPvm neurons. (A) Each VPdl neuron’s directional change in FR during the approach (y-axis) plotted against its directional change in FR during the response (x-axis). Each dot represents one neuron. Two example VPdl neurons exhibiting similar directional changes in FR from approach to response are displayed in (B) and (C). (D) Each VPvm neuron’s directional change in FR during the approach (y-axis) plotted against its directional change in FR during the response (x-axis). Each dot represents one neuron. One example VPvm neuron exhibiting similar directional changes in FR from approach to response is displayed in (E). Rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots in approach-rasters and blue dots in response-rasters indicate the onset of approach and onset of response, respectively. Red dots in retreat-rasters indicate the offset of retreat. All rasters have been sorted for
movement duration. Y-axis of raster refers to behavioral trial; y-axis of PETH refers to average FR (spikes/sec) per trial. Both x-axes refer to time (ms).

Figure 11. Correlations for other changes in FR during two behaviors, Approach-Retreat or Response-Retreat, did not differ between subregions. Each VPdl (A) and VPvm (B) neuron’s, directional change in FR during the approach (y-axis) was plotted against its directional change in FR during the retreat (x-axis). Each dot represents one neuron. Two examples of changes in FR during approach and retreat, but not during response, are displayed in (C) and (D), both VPvm neurons. To examine response-retreat relationships, each VPdl (E) and VPvm (F) neuron’s directional change in FR during the response (y-axis) was plotted against its directional change in FR during the retreat (x-axis). Each dot represents one neuron. Two examples of changes in FR during response and retreat, but not during approach, are displayed ((G) for VPdl neuron and (H) for VPvm neuron). For (C), (D), (G), and (H), rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots indicate onset of approach and blue dots indicate onset of response. Red dots indicate offset of retreat. All rasters have been sorted for movement duration. Y-axes of rasters refer to behavioral trial; y-axes of PETHs refer to average FR (spikes/sec) per trial. Both x-axes refer to time (ms).
Figure 12. Changes in FR during all three behaviors, Approach-Response-Retreat. Some neurons exhibited continuous directional changes in FR starting at the approach and continuing through the response and retreat. Two examples of similar directional changes in FR between approach, response, and retreat changes in FR, one VPdl neuron (A) and one VPvm neuron (B) are displayed. Many neurons exhibited heterogeneous directional changes in FR between approach, response, and retreat changes in FR. Six examples of heterogeneous directional changes in FR between approach, response, and retreat changes in FR, two VPdl neurons (C-D) and four VPvm neurons (E-H), are displayed. For A-H, rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots indicate onset of approach and blue dots indicate onset of response. Red dots indicate the offset of retreat. All rasters have been sorted for movement duration. Y:axis of raster refers to behavioral trial; y:axis of PETH refers to average FR (spikes/sec) per trial. Both x:axes refer to time (ms).

Figure 13. Absence of "initiation" firing patterns by VP neurons in the present task. Three candidate neurons are displayed which were the only neurons that, upon first inspection exhibited a potential firing pattern prior to the onset of approach (green dots in left panels of A-C). Blue dots and red dots in A-C indicate onset of response and offset of retreat, respectively. Time 0 (ms) of A-C (left) indicates offset of approach, time 0 (ms) of A-C (second from left) indicates offset of response, and time 0 (ms) of A-C (third from left) indicates onset of
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retreat. A (second from right) and B (second from right) are the same data from A (left) and B (left), respectively, except in A (second from right) and B (second from right) the onset of approach was redisplayed at time 0 and the offset of approach was displayed as green dots. Video analyses revealed that these neurons, recorded from the same animal, changed FRs during a locomotor movement prior to the approach rather than exhibiting an "initiation" (pre-movement) firing pattern (A (second from right), B (second from right), also see Movie 2). In A (right) and B (right) magenta dots indicate the onset and cyan dots indicate offset of the pre-approach movement. For neuron in (C), the onset of approach was redisplayed at time 0 (ms) and offset of approach was indicated by green dots (first, second, and third panels from right). We examined whether changes in FR prior to the approach in C (left), were the result of approaches that were preceded within close proximity to other behaviors. C (third from right) displays trials in which a previous approach occurred within the 4 second firing window prior to the current approach. Prior approach onset and offset are displayed as magenta and cyan dots, respectively. C (second from right) displays trials in which a previous response occurred within the 4 second firing window prior to the approach. Prior response onset and offset are displayed as magenta and cyan dots, respectively. C (right) displays trials in which a previous retreat occurred within the 4 second firing window prior to the approach. Prior retreat onset and offset are displayed as magenta and cyan dots, respectively. This figure indicates that the decrease in FR observed in C (left) prior to the approach was due to a decrease in FR near the offset of the previous retreat which was
proximal to the approach onset (C (right)). All rasters have been sorted for movement duration. Y-axis of raster refers to behavioral trial; y-axis of PETH refers to average FR (spikes/sec) per trial. Both x-axes refer to time (ms).
Table 1. Primary Antibodies Used

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<th>Antigen (what is being stained for)</th>
<th>Immunogen (what the antibody was raised against; full sequence and species)</th>
<th>Manufacturer, species antibody was raised in, mono- vs. polyclonal, catalog or lot number</th>
<th>Dilution used</th>
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Table 2. Electrophysiological data by subregion

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<th>Subregion</th>
<th>Units (n)</th>
<th>Triphasic (%)</th>
<th>Amplitude (µV)</th>
<th>Signal-to-noise ratio</th>
<th>Baseline FR (cue)</th>
<th>Baseline FR (behaviors)</th>
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<td>VPdl</td>
<td>48</td>
<td>8.33</td>
<td>116.55 ± 7.10</td>
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<td>11.63</td>
<td>139.14 ± 6.33</td>
<td>4.22 ± 0.09</td>
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<td>Total</td>
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<td>Behavior</td>
<td>Number of events per animal</td>
<td>Duration (ms)</td>
<td></td>
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<tr>
<td>Approach</td>
<td>608.96 ± 72.77</td>
<td>956.60 ± 63.08</td>
<td></td>
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<td></td>
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<tr>
<td>Cued Approach</td>
<td>310.60 ± 35.17</td>
<td>955.97 ± 63.83</td>
<td></td>
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<tr>
<td>Uncued Approach</td>
<td>298.36 ± 38.98</td>
<td>960.40 ± 62.55</td>
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<tr>
<td>Criterion Movement (Response)</td>
<td>312.96 ± 41.72</td>
<td>340.20 ± 19.00</td>
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<tr>
<td>Cued Response</td>
<td>147.52 ± 18.82</td>
<td>341.08 ± 18.88</td>
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<tr>
<td>Uncued Response</td>
<td>165.44 ± 24.44</td>
<td>335.95 ± 20.52</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Retreat</td>
<td>630.00 ± 75.09</td>
<td>694.92 ± 39.00</td>
<td></td>
<td></td>
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</tr>
</tbody>
</table>
Table 4. Distribution of neurons within VP subregions exhibiting 20% change in firing rate from baseline (increase, decrease, or mixed) during distinct behaviors.

<table>
<thead>
<tr>
<th></th>
<th>Increase (% / N)</th>
<th>Decrease (% / N)</th>
<th>Mixed (% / N)</th>
<th>Total (% / N)</th>
</tr>
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<tbody>
<tr>
<td>Approach VPdl</td>
<td>4.17% / 2</td>
<td>6.25% / 3</td>
<td>NA</td>
<td>10.42% / 5</td>
</tr>
<tr>
<td>Approach VPvm</td>
<td>2.92% / 5</td>
<td>5.26% / 9</td>
<td>NA</td>
<td>8.19% / 14</td>
</tr>
<tr>
<td>Response VPdl</td>
<td>6.25% / 3</td>
<td>18.75% / 9</td>
<td>NA</td>
<td>25% / 12</td>
</tr>
<tr>
<td>Response VPvm</td>
<td>10.53% / 18</td>
<td>12.28% / 21</td>
<td>NA</td>
<td>22.81% / 39</td>
</tr>
<tr>
<td>Retreat VPdl</td>
<td>0% / 0</td>
<td>2.08% / 1</td>
<td>NA</td>
<td>2.08% / 1</td>
</tr>
<tr>
<td>Retreat VPvm</td>
<td>5.85% / 10</td>
<td>2.92% / 5</td>
<td>NA</td>
<td>8.77% / 15</td>
</tr>
<tr>
<td>Approach-Response VPdl</td>
<td>2.08% / 1</td>
<td>10.42% / 5</td>
<td>2.08% / 1</td>
<td>14.58% / 7</td>
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<tr>
<td>Approach-Response VPvm</td>
<td>1.75% / 3</td>
<td>5.85% / 10</td>
<td>1.75% / 3</td>
<td>9.36% / 16</td>
</tr>
<tr>
<td>Approach-Retreat VPdl</td>
<td>4.17% / 2</td>
<td>2.08% / 1</td>
<td>0% / 0</td>
<td>6.25% / 3</td>
</tr>
<tr>
<td>Approach-Retreat VPvm</td>
<td>2.34% / 4</td>
<td>2.34% / 4</td>
<td>1.17% / 2</td>
<td>5.85% / 10</td>
</tr>
<tr>
<td>Response-Retreat VPdl</td>
<td>0% / 0</td>
<td>2.08% / 1</td>
<td>2.08% / 1</td>
<td>4.17% / 2</td>
</tr>
<tr>
<td>Response-Retreat VPvm</td>
<td>1.75% / 3</td>
<td>2.34% / 4</td>
<td>7.02% / 12</td>
<td>11.11% / 19</td>
</tr>
<tr>
<td>Approach-Response-Retreat VPdl</td>
<td>6.25% / 3</td>
<td>12.5% / 6</td>
<td>4.17% / 2</td>
<td>22.92% / 11</td>
</tr>
<tr>
<td>Approach-Response-Retreat VPvm</td>
<td>2.92% / 5</td>
<td>3.51% / 6</td>
<td>3.51% / 6</td>
<td>9.94% / 17</td>
</tr>
<tr>
<td>Total VPdl (% / N)</td>
<td>22.92% / 11</td>
<td>54.17% / 26</td>
<td>8.33% / 4</td>
<td>85.42% / 41</td>
</tr>
<tr>
<td>Total VPvm (% / N)</td>
<td>28.07% / 48</td>
<td>34.5% / 59</td>
<td>13.45% / 23</td>
<td>76.02% / 130</td>
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Figure 1. Behaviors schematized for neuronal analysis. Appetitive behaviors included the approach towards the photocell corner (A), criterion head movement operant response (B), and retreat away from the photocell corner (C). Arrows in (A) and (C) indicate locomotion that is absent from (B). The criterion movement consisted of a vertical head movement breaking photocells 1 or 2 through 5 or 6, consecutively, under one second in duration. In this example, the rat executes a vertical head movement breaking photocells 1 through 6.
Figure 2. Self-administration and cue learning. Reaction time in response to the SD (A), probability of self-administering cocaine upon SD presentation (B), cocaine consumption (C), latency to load up to the tenth infusion (D), number of criterion movements (E). Values are average ± SEM (y axes) per day (x axes).
Figure 3. Skill learning: changes in criterion head movement parameters with extensive repetition. Changes in velocity (A), duration (B), distance (C), probability of starting at photocell two, the minimum required for a criterion movement (D), and the probability of ending at photocell 5 (E). Values for A, B, and C are average median ± SEM per day for criterion head movements (y axes). Values for D and E are average probability ± SEM. All x axes refer to training day.
Figure 4. Example microwires localized to VP subregions. Examples are displayed in groups of three. A-I displays three examples of microwires localized to the VPdl in three vertical panels each; example one A-C, example two D-F, and example three G-I. J-R display three examples of microwires localized to the VPvm in three vertical panels each; example one J-L, example two M-O, and example three P-R. Panels A, D, G, J, M, and P display substance P immunohistochemistry. Panels B, E, H, K, N, and Q display calbindin-d28k immunohistochemistry. Panels C, F, I, L, O, and R display neurotensin immunohistochemistry. Green/blue dot in each panel is an iron deposit from the uninsulated microwire tip visualized from by potassium ferrocyanide counterstain. Numbers refer to approximate anteroposterior coordinate based on Paxinos and Watson (2004). As a model for all VPdl neurons recorded, VPdl example 1 (A-C) displays two microwire tip locations. The wire closest to the anterior commissure was localized to substance P immunoreactivity (A) and calbindin-d28k immunoreactivity (B), but not neurotensin immunoreactivity (C). The microwire furthest from the anterior commissure (A) was localized outside of substance P immunoreactivity and therefore excluded from the dataset. As a model for all VPvm neurons recorded, VPvm example 1 (J-L) displays a
microwire tip location localized to substance P immunoreactivity (J) and neurotensin immunoreactivity (K), but not calbindin-d28k immunoreactivity (L). Any tissue from the left hemisphere has been rotated horizontally to the right hemisphere for the sake of uniform orientation (midline is left for all panels). Outlines were based on substance P, calbindin-d28k, and neurotensin immunoreactivity, the atlas of Paxinos and Watson (2004), and scientific literature (Zahm and Heimer, 1988, 1990; Zahm 1989; Zahm et al. 1996; Riedel et al. 2002; Tripathi et al. 2010). Calibration bar in A represents 1 mm. All sections were 40 µm thick.

159x256mm (300 x 300 DPI)
Figure 5. All microwires from all rats that were localized to VP subregions. Tissue displayed is substance P immunoreactivity from the left hemisphere of a rat used in the present dataset. The left hemisphere implant from this rat was misplaced dorsal to VP and thus unencumbered by microwire tip counterstaining, and therefore was selected for display purposes. Black circles and cyan triangles indicate VPvm and VPDl localized microwires, respectively. A-J display right hemisphere localized microwires (midline right). K-U display left hemisphere localized microwires (midline left). Outlines were based on substance P immunoreactivity as well as the atlas of Paxinos and Watson (2004). Numbers refer to approximate anteroposterior coordinate based on Paxinos and Watson (2004). Calibration bar in A represents 1 mm. All sections were 40 μm thick.

155x177mm (300 x 300 DPI)
Figure 6. VPdl neurons exhibit greater changes in FR during the approach than VPvm neurons whether cued or uncued. (A) Cumulative proportion of all neurons' absolute changes in FR during the approach per subregion. Gray and black lines indicate cumulative proportions for VPdl and VPvm neurons, respectively. Median values (0.50 proportion value) are displayed as circle (VPdl) and square (VPvm). For this and all other cumulative proportion figures, the absolute changes in FR were used. A value of 0 was no difference from baseline FR and a fifty percent change from baseline FR was 0.1. Examples of approach-related changes in FR are displayed in (B) from a VPdl neuron and (C) from a VPvm neuron. For (B) and (C), rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots in approach-raster and blue dots in response-raster indicate the onset of approach and onset of response, respectively. Red dots in retreat-raster indicate the offset of retreat. The cumulative proportions of all neurons' absolute changes in FR for cued approach and uncued approach per subregion are shown in (D) and (E), respectively. Each VPdl (F) and VPvm neuron (G) directional change in FR during the cued approach (y-axis) was plotted against its directional change in FR during the uncued approach.
approach (x-axis). Each dot represents one neuron. Note that for (F) and (G), and all other scatter plot figures, directional changes in FR were used. In contrast to the absolute change in FR, a value of 0.5 was not different from baseline FR and a fifty percent change from baseline FR was 0.6 (increase) or 0.4 (decrease). Two examples of similar directional changes in FR between cued and uncued approach changes in FR, one VPdl neuron (H) and one VPvm neuron (I) are displayed. For (H) and (I), rasters and PETHs are centered (time 0) around offset of cued approach (left) and uncued approach (right). Green dots indicate the onset of approach. All rasters have been sorted for movement duration. Y-axes of all rasters refer to behavioral trial; y-axes of all PETHs refer to average FR (spikes/sec) per trial. All raster and PETH x-axes refer to time (ms). *** p < 0.001, ** p < 0.01; * p < 0.05.

201x286mm (300 x 300 DPI)
Figure 7. VPdl neurons exhibit greater changes in FR during the response than VPvm neurons whether cued or uncued. (A) Cumulative proportion of all neurons' absolute changes in FR during the response per subregion. Gray and black lines indicate cumulative proportions for VPdl and VPvm neurons, respectively. Median values (0.50 proportion value) are displayed as circle (VPdl) and square (VPvm). Examples of approach-related changes in FR are displayed in (B) from a VPdl neuron and (C) from a VPvm neuron. For (B) and (C), rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots in approach-raster and blue dots in response-raster indicate the onset of approach and onset of response, respectively. Red dots in retreat-raster indicate the offset of retreat. The cumulative proportion of all neurons' absolute changes in FR for cued approach and uncued approach per subregion are shown in (D) and (E), respectively. Each VPdl (F) and VPvm neuron (G) directional change in FR during the cued approach (y-axis) was plotted against its directional change in FR during the uncued approach (x-axis). Each dot represents one neuron. Two examples of similar directional changes in FR between cued and uncued approach changes in FR, one VPdl neuron (H) and one VPvm
neuron (I) are displayed. For (H) and (I), rasters and PETHs are centered (time 0) around offset of cued response (left) and uncued response (right). Blue dots indicate the onset of response. All rasters have been sorted for movement duration. Y-axes of all rasters refer to behavioral trial; y-axes of all PETHs refer to average FR (spikes/sec) per trial. All raster and PETH x-axes refer to time (ms). ** p < 0.01; * p < 0.05.
162x238mm (300 x 300 DPI)
Figure 8. No relationship between response movement parameters and changes in FR by VP neurons. Given that no significant relationships were observed, all neurons were pooled into graphs of distance (A), duration (B), velocity (C), valley (start position) (D), and peak (end position) (E) regardless of subregion. Each bin on the x-axis is 0.10. Y-axis is frequency of occurrence (number of neurons exhibiting the r value).
Figure 9. No differences between subregions during the retreat. Cumulative proportion of all neurons' absolute change in FR during the retreat per subregion. Gray and black lines in (A) indicate cumulative proportions for VPdl and VPvm neurons, respectively. Median values (0.50 proportion value) are displayed as circle (VPdl) and square (VPvm) in (A). One example of retreat-related firing is displayed in (B), from a VPvm neuron. Raster and PETH are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dot in approach-raster and blue dot in response-raster indicate the onset of approach and response, respectively. Red dot in C indicates the offset of retreat. For each VPdl (C) and VPvm neuron (D), the neuron's directional change in FR during the nonpump retreat (y-axis) was plotted against its directional change in FR during the pump retreat (x-axis). Each dot represents one neuron. Two examples of similar directional changes in FR between pump retreat and nonpump retreat changes in FR, one VPdl neuron (E) and one VPvm neuron (F) are displayed. Two examples of dissimilar directional changes in FR between pump retreat and nonpump retreat changes in FR, one VPdl neuron (G) and one VPvm neuron (H) are displayed. For (E), (F), (G), and (H), rasters and PETHs are centered (time 0)
around onset of pump retreats (left) and nonpump retreats (right). Red dots indicate the offset of the retreat. All rasters have been sorted for movement duration. Y-axes of rasters refer to behavioral trial; y-axes of PETHs refer to average FR (spikes/sec) per trial. Both x-axes refer to time (ms).
Figure 10. VPdl neurons exhibit a greater correlation between approach-and-response changes in FR than VPvm neurons. (A) Each VPdl neuron’s directional change in FR during the approach (y-axis) plotted against its directional change in FR during the response (x-axis). Each dot represents one neuron. Two example VPdl neurons exhibiting similar directional changes in FR from approach to response are displayed in (B) and (C). (D) Each VPvm neuron’s directional change in FR during the approach (y-axis) plotted against its directional change in FR during the response (x-axis). Each dot represents one neuron. One example VPvm neuron exhibiting similar directional changes in FR from approach to response is displayed in (E). Rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots in approach-rasters and blue dots in response-rasters indicate the onset of approach and onset of response, respectively. Red dots in retreat-rasters indicate the offset of retreat. All rasters have been sorted for movement duration. Y-axis of raster refers to behavioral trial; y-axis of PETH refers to average FR (spikes/sec) per trial. Both x-axes refer to time (ms).
Figure 11. Correlations for other changes in FR during two behaviors, Approach-Retreat or Response-Retreat, did not differ between subregions. Each VPdl (A) and VPvm (B) neuron’s, directional change in FR during the approach (y-axis) was plotted against its directional change in FR during the retreat (x-axis). Each dot represents one neuron. Two examples of changes in FR during approach and retreat, but not during response, are displayed in (C) and (D), both VPvm neurons. To examine response-retreat relationships, each VPdl (E) and VPvm (F) neuron’s directional change in FR during the response (y-axis) was plotted against its directional change in FR during the retreat (x-axis). Each dot represents one neuron. Two examples of changes in FR during response and retreat, but not during approach, are displayed ((G) for VPdl neuron and (H) for VPvm neuron). For (C), (D), (G), and (H), rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots indicate onset of approach and blue dots indicate onset of response. Red dots indicate offset of retreat. All rasters have been sorted for movement duration. Y-axes of rasters refer to behavioral trial; y-axes of PETHs refer to average FR (spikes/sec) per trial. Both x-axes refer to time (ms).
Figure 12. Changes in FR during all three behaviors, Approach-Response-Retreat. Some neurons exhibited continuous directional changes in FR starting at the approach and continuing through the response and retreat. Two examples of similar directional changes in FR between approach, response, and retreat changes in FR, one VPdl neuron (A) and one VPvm neuron (B) are displayed. Many neurons exhibited heterogeneous directional changes in FR between approach, response, and retreat changes in FR. Six examples of heterogeneous directional changes in FR between approach, response, and retreat changes in FR, two VPdl neurons (C-D) and four VPvm neurons (E-H), are displayed. For A-H, rasters and PETHs are centered (time 0) around offset of approach (left), offset of response (center), and onset of retreat (right). Green dots indicate onset of approach and blue dots indicate onset of response. Red dots indicate the offset of retreat. All rasters have been sorted for movement duration. Y-axis of raster refers to behavioral trial; y-axis of PETH refers to average FR (spikes/sec) per trial. Both x-axes refer to time (ms).
Figure 13. Absence of "initiation" firing patterns by VP neurons in the present task. Three candidate neurons are displayed which were the only neurons that, upon first inspection exhibited a potential firing pattern prior to the onset of approach (green dots in left panels of A-C). Blue dots and red dots in A-C indicate onset of response and offset of retreat, respectively. Time 0 (ms) of A-C (left) indicates offset of approach, time 0 (ms) of A-C (second from left) indicates offset of response, and time 0 (ms) of A-C (third from left) indicates onset of retreat. A (second from right) and B (second from right) are the same data from A (left) and B (left), respectively, except in A (second from right) and B (second from right) the onset of approach was redisplayed at time 0 and the offset of approach was displayed as green dots. Video analyses revealed that these neurons, recorded from the same animal, changed FRs during a locomotor movement prior to the approach rather than exhibiting an "initiation" (pre-movement) firing pattern (A (second from right), B (second from right), also see Movie 2). In A (right) and B (right) magenta dots indicate the onset and cyan dots indicate offset of the pre-approach movement. For neuron in (C), the onset of approach was redisplayed at time 0 (ms) and offset of approach was indicated by green dots (first, second, and third
panels from right). We examined whether changes in FR prior to the approach in C (left), were the result of approaches that were preceded within close proximity to other behaviors. C (third from right) displays trials in which a previous approach occurred within the 4 second firing window prior to the current approach. Prior approach onset and offset are displayed as magenta and cyan dots, respectively. C (second from right) displays trials in which a previous response occurred within the 4 second firing window prior to the approach. Prior response onset and offset are displayed as magenta and cyan dots, respectively. C (right) displays trials in which a previous retreat occurred within the 4 second firing window prior to the approach. Prior retreat onset and offset are displayed as magenta and cyan dots, respectively. This figure indicates that the decrease in FR observed in C (left) prior to the approach was due to a decrease in FR near the offset of the previous retreat which was proximal to the approach onset (C (right)). All rasters have been sorted for movement duration. Y-axis of raster refers to behavioral trial; y-axis of PETH refers to average FR (spikes/sec) per trial. Both x-axes refer to time (ms).
MOVIE LEGENDS

Movie 1. This movie provides four examples of approach, response, and retreat events used for electrophysiological analysis. Onset and offset times are marked for each behavior. Approach onset, retreat onset, and retreat offset were determined using frame-by-frame video scoring. Note that approach offset, response onset, and response offset were determined via electronic timestamp of photocell beam breaks. These timepoints were displayed in this video for illustrative purposes only.

Movie 2. This movie provides examples of the pre-approach movement and approach events exhibited by one rat. Two neurons exhibited changes in FR during these behaviors (Figure 13A,B). Onset and offset times are marked for both behaviors.